

Biomaterials dedicated to bone regeneration

**SNOSCELLS
Les Houches**

Pierre Weiss

INSERM U 1229

RMES Regenerative medicine and skeleton,

University of Nantes, 1 place Alexis Ricordeau, 44042 Nantes,
France.

E-mail : pierre.weiss@univ-nantes.fr

Day 2

Regenerative biomaterials for bone

- Biomaterials in bone regenerative medicine.

Biomaterials for bone regeneration

❑ Autogenous bone (Gold Standard)

- Limited bone stock (small packs)
- High risk of morbidity
- Painful and not aesthetic

❑ Allograft (Human)

❑ Xenograft bone (Animal)

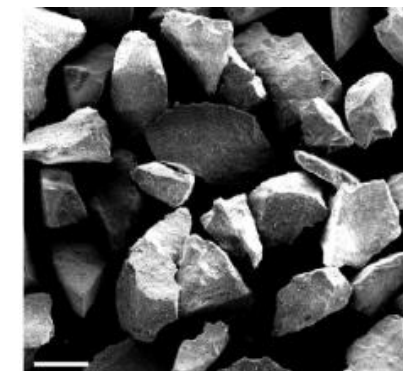
- Infection (bacterial - viral)
- Dead bones limit cell proliferation
- Legal requirements (biological origins)

❑ Calcium Phosphates

- Wide variety on the market
- Different quality and clinical results
- Very specific production know-how

❑ Bio Glasses

- Controlled synthetic material
- Able to recruit osteoprogenitor cells, the "stem cells" of bone.
- controls the cycle of osteoblasts to promote proliferation and differentiation



Biomaterial Design

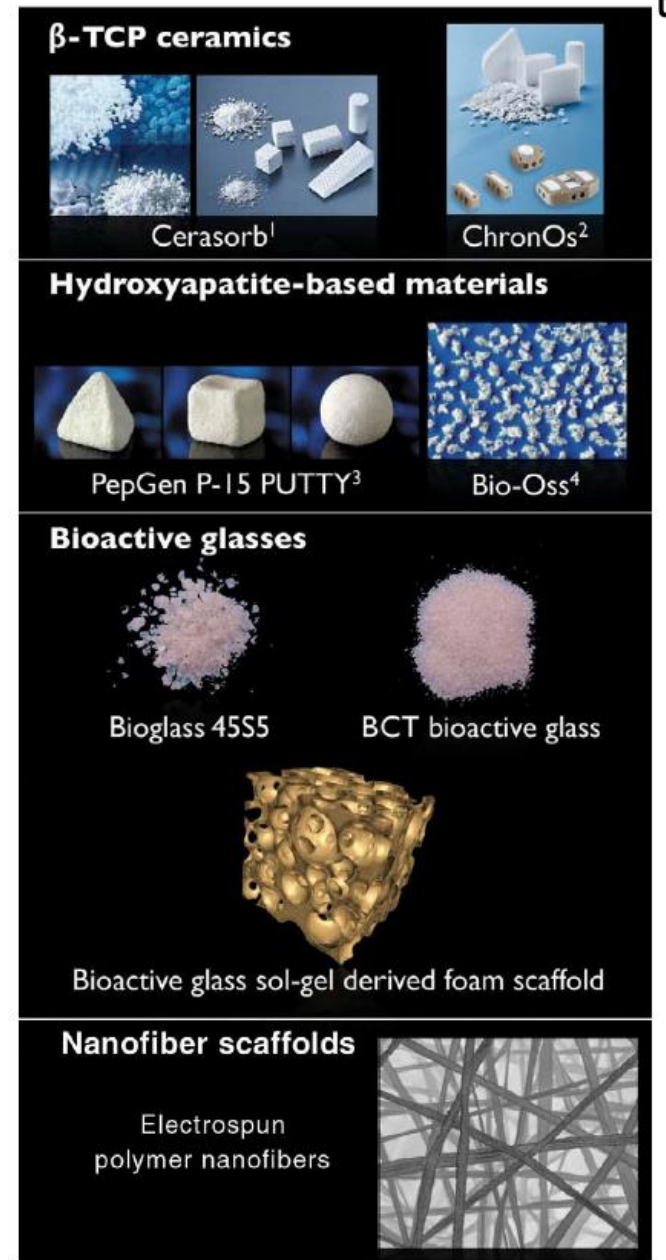
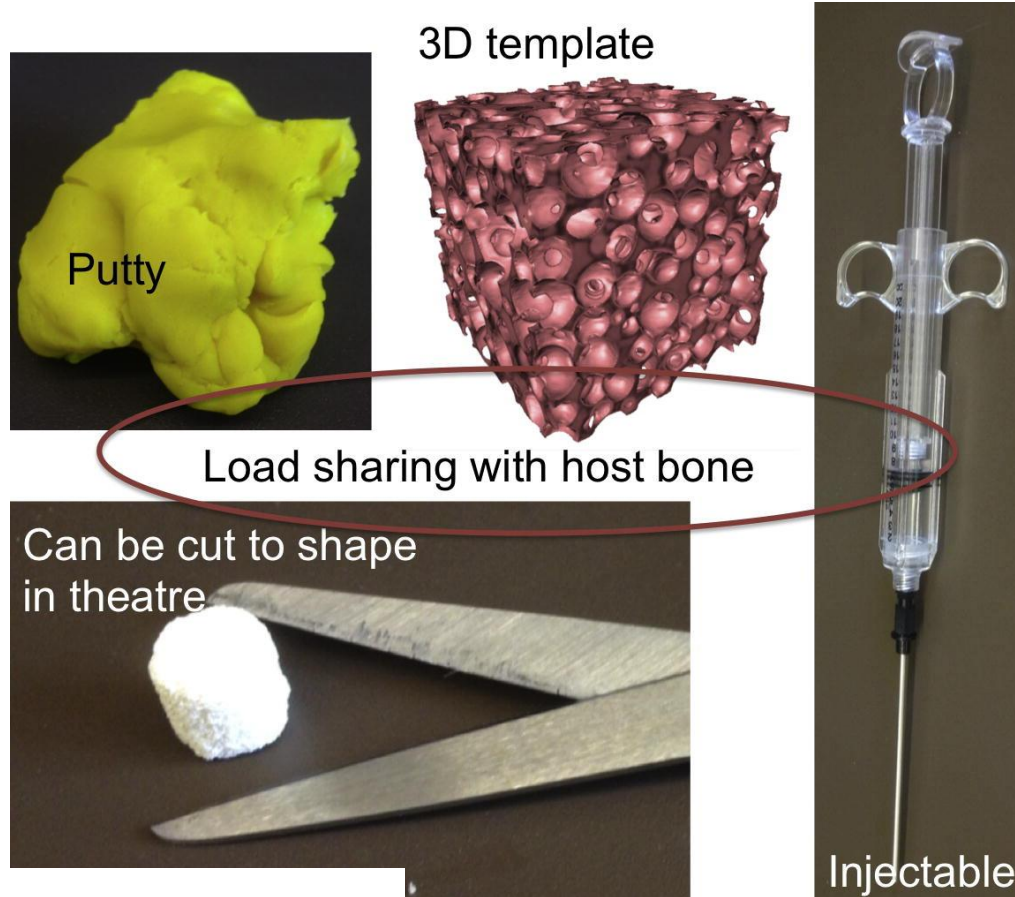


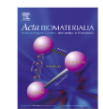
Fig. 2. Macromorphology of some examples of different bone graft materials. (Reproduced with permission from ¹Curasan AG, ²Synthes, ⁴Geistlich. ³Courtesy of Dentsply Tulsa Dental Specialties. © PepGen® P-15.)

Acta Biomaterialia 23 (2015) 553–582

Contents lists available at ScienceDirect

Acta Biomaterialia

journal homepage: www.elsevier.com/locate/actabiomat



Review

Reprint of: Review of bioactive glass: From Hench to hybrids

Julian R. Jones*

Department of Materials, Imperial College London, South Kensington Campus, London SW7 2AZ, UK



Calcium Phosphate pionniers



Biological and synthetic apatites

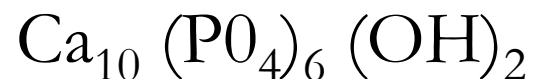
Bone mineral:



Dental enamel Mineral:



Hydroxyapatite

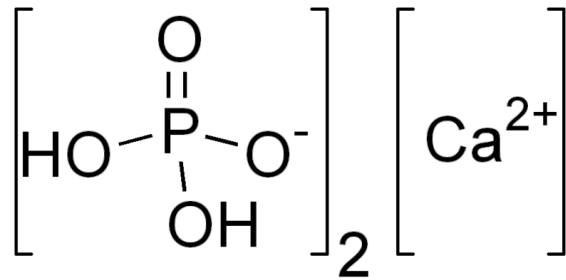


Beta Tricalcique Phosphate (β -TCP) :

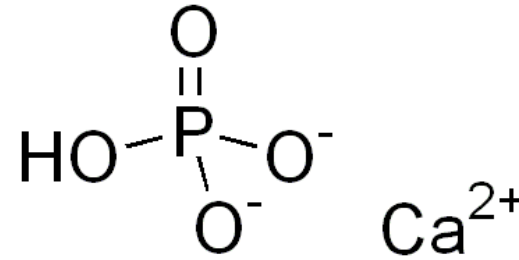


Biphasic calcium phosphate BCP = HA + β -TCP

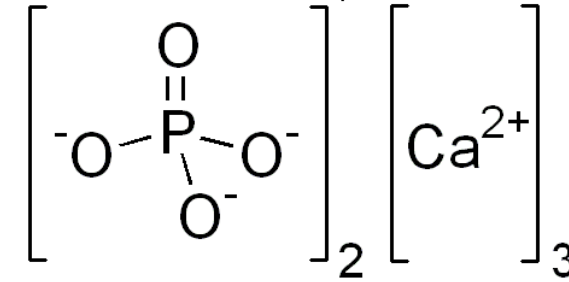
Monocalcium Phosphate



Dicalcium phosphate



tricalcium Phosphate : TCP :



1468

S.V. Dorozhkin / Biomaterials 31 (2010) 1465–1485

Table 1

Existing calcium orthophosphates and their major properties [21,22].

Ca/P molar ratio	Compound	Formula	Solubility at 25 °C, $-\log(K_s)$	Solubility at 25 °C, g/L	pH stability range in aqueous solutions at 25 °C
0.5	Monocalcium phosphate monohydrate (MCPM)	$\text{Ca}(\text{H}_2\text{PO}_4)_2 \cdot \text{H}_2\text{O}$	1.14	~ 18	0.0–2.0
0.5	Monocalcium phosphate anhydrous (MCPA)	$\text{Ca}(\text{H}_2\text{PO}_4)_2$	1.14	~ 17	^c
1.0	Dicalcium phosphate dihydrate (DCPD), mineral brushite	$\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$	6.59	~ 0.088	2.0–6.0
1.0	Dicalcium phosphate anhydrous (DCPA), mineral monetite	CaHPO_4	6.90	~ 0.048	^c
1.33	Octacalcium phosphate (OCP)	$\text{Ca}_8(\text{HPO}_4)_2(\text{PO}_4)_4 \cdot 5\text{H}_2\text{O}$	96.6	~ 0.0081	5.5–7.0
1.5	α -Tricalcium phosphate (α -TCP)	$\alpha\text{-Ca}_3(\text{PO}_4)_2$	25.5	~ 0.0025	^a
1.5	β -Tricalcium phosphate (β -TCP)	$\beta\text{-Ca}_3(\text{PO}_4)_2$	28.9	~ 0.0005	^a
1.2–2.2	Amorphous calcium phosphate (ACP)	$\text{Ca}_x\text{H}_y(\text{PO}_4)_z \cdot n\text{H}_2\text{O}$, $n = 3\text{--}4.5$; 15–20% H_2O	^b	^b	~ 5–12 ^d
1.5–1.67	Calcium-deficient hydroxyapatite (CDHA) ^e	$\text{Ca}_{10-x}(\text{HPO}_4)_x(\text{PO}_4)_{6-x}(\text{OH})_{2-x}$ ($0 < x < 1$)	~ 85.1	~ 0.0094	6.5–9.5
1.67	Hydroxyapatite (HA or OHAp)	$\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$	116.8	~ 0.0003	9.5–12
1.67	Fluorapatite (FA or FAp)	$\text{Ca}_{10}(\text{PO}_4)_6\text{F}_2$	120.0	~ 0.0002	7–12
2.0	Tetracalcium phosphate (TTCP or TetCP), mineral hilgenstockite	$\text{Ca}_4(\text{PO}_4)_2\text{O}$	38–44	~ 0.0007	^a

CA

^a These compounds cannot be precipitated from aqueous solutions.

^b Cannot be measured precisely. However, the following values were found: 25.7 ± 0.1 (pH = 7.40), 29.9 ± 0.1 (pH = 6.00), 32.7 ± 0.1 (pH = 5.28). The comparative extent of dissolution in acidic buffer is: ACP >> α -TCP >> β -TCP > CDHA >> HA > FA.

^c Stable at temperatures above 100 °C.

^d Always metastable.

^e Occasionally, CDHA is named as precipitated HA.

^f In the case $x = 1$ (the boundary condition with Ca/P = 1.5), the chemical formula of CDHA looks as follows: $\text{Ca}_9(\text{HPO}_4)(\text{PO}_4)_5(\text{OH})$.

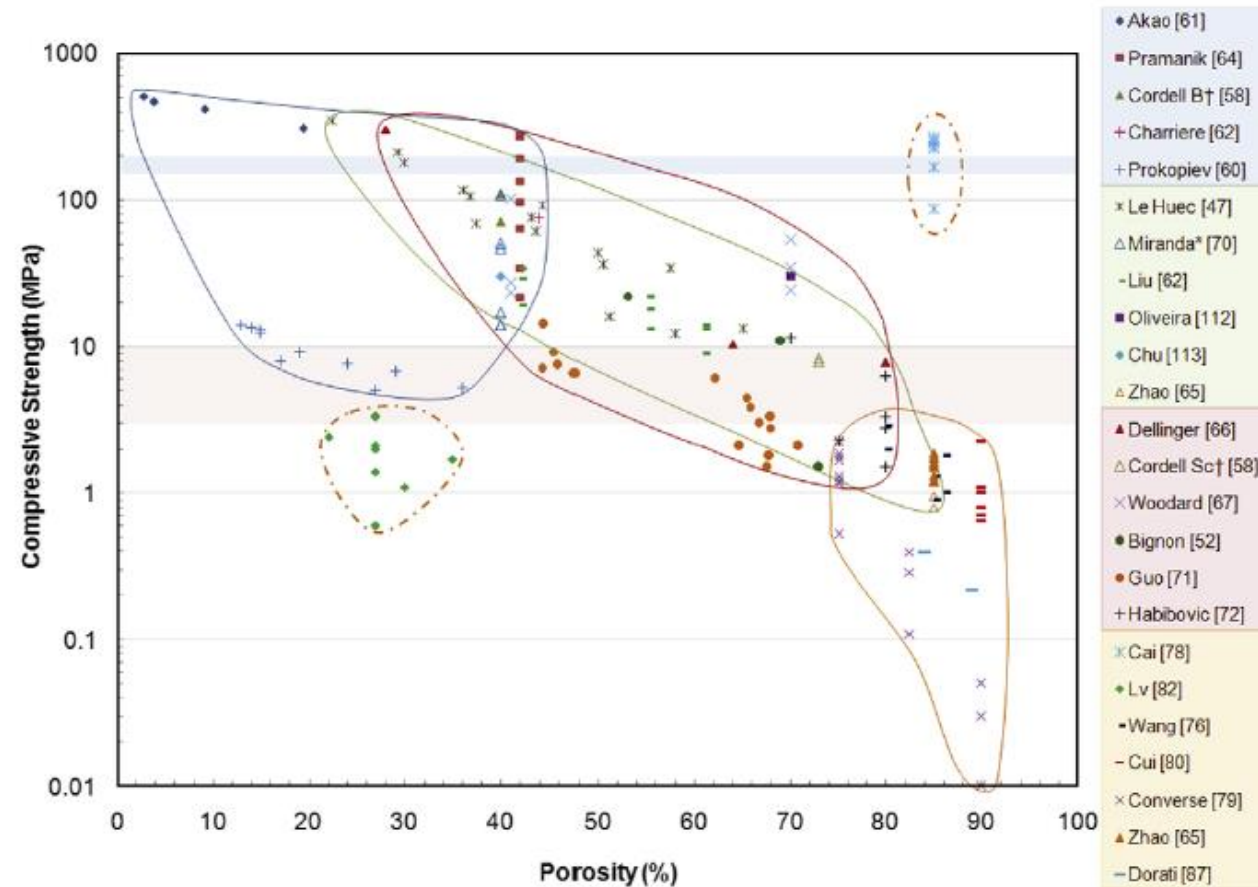


Fig. 2. Plot illustrating the dependence of compressive strength on total porosity for all references included in Fig. 1. The data are grouped by material category using colored lines. Bulk HA and BCP, HA scaffolds, BCP scaffolds, HA/polymer and polymer scaffolds are represented by blue, green, red and orange lines, respectively. One exception is Miranda et al. (denoted by *). These data are included in the BCP category, but are the only study in that category with β -TCP and no HA. Two data sets from two different groups are considered "outliers" and are demarcated with dashed lines. Table 2 is also the legend to this figure.

Biological activity of CaP

J. Van der Stok et al./Acta Biomaterialia 7 (2011) 739–750

743

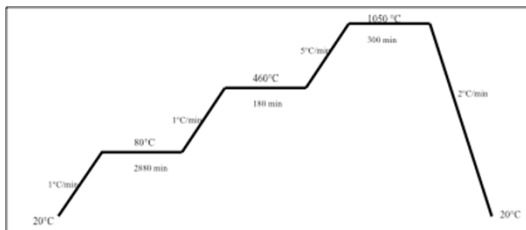
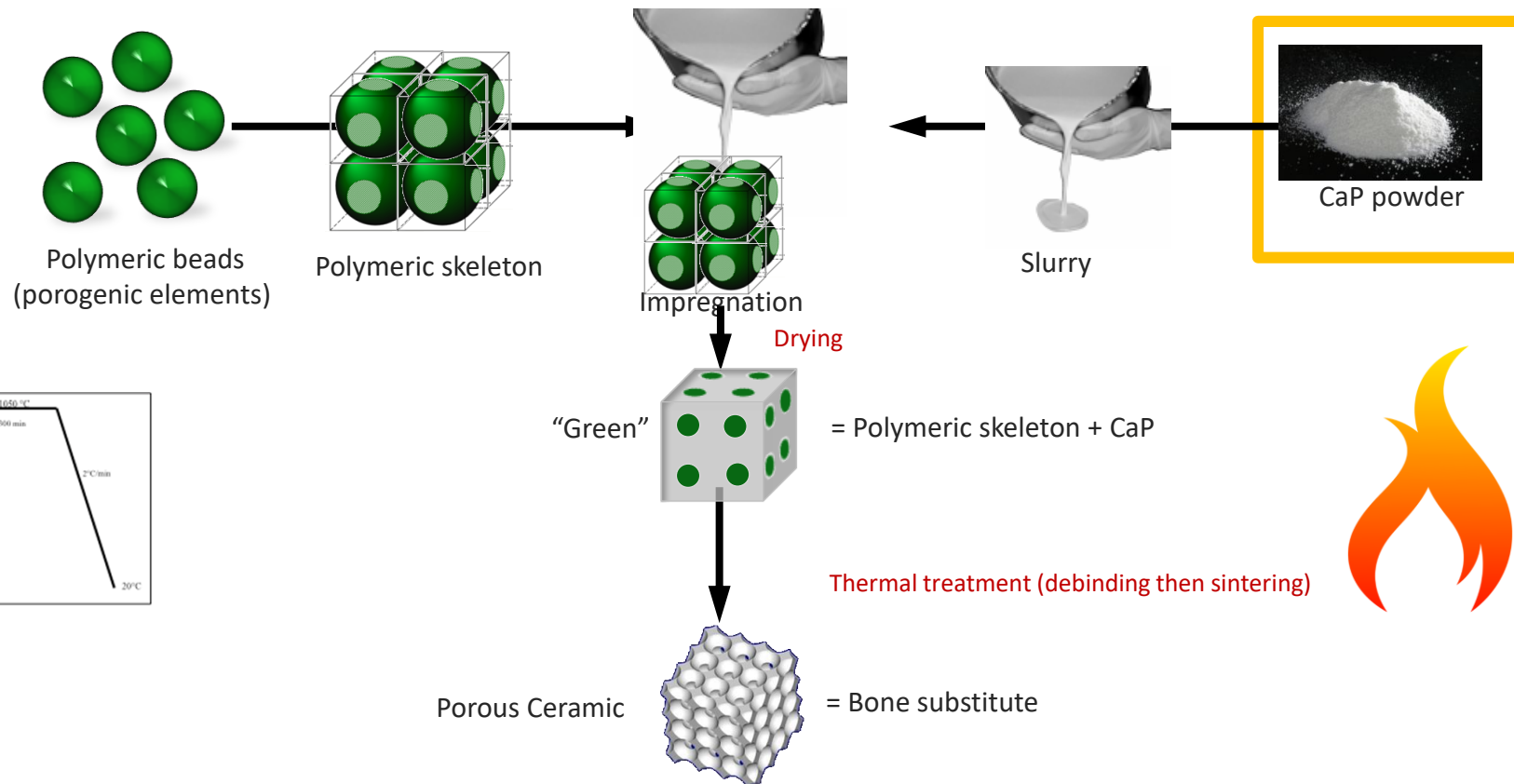
Table 3
Overview of porosity, pore size and biological properties.

Product name	Porosity (%)	Pore size (µm)	Osteogenic	Osteoinductive	Osteoconductive	Biodegradable
<i>Calcium phosphate</i>						
<i>Hydroxyapatite</i>						
Cerabone®	N.D.	(100–1500)	N.D.	N.D.	Yes [47]	N.D.
Endobon®	60–80 [24]	390–1360 [22]	N.D.	No [38]	Yes [24,38,41,43,44,65,82,83,159,160]	No [39–44]
Ostim®	N.D.	N.D.	N.D.	N.D.	Yes [45–50,85,161,162]	Yes [45–50]
Pro Osteon 500®	N.D.	(280–770)	N.D.	N.D.	Yes [40,163,164]	No [165]
<i>Tricalcium phosphate</i>						
ChronOS™	60–75 [25,27,28]	100–400 [25,28]	N.D.	N.D.	Yes [25,27,166–168]	Yes [27,167,169]
Vitoss®	(88–92)	(1–1000)	N.D.	N.D.	Yes [25,170]	Yes [25]
<i>Composite</i>						
BoneSave®	(50)	(300–500)	N.D.	N.D.	N.D.	N.D.
BoneSource®	46 [29,30]	2–50 [31]	N.D.	Yes [29]	Yes [30,72,74,110,171–176]	Yes [72,171,172,174,175]
Calcibon®	30–40 [32,33]	<1 [33]	N.D.	No [37]	Yes [33,37,57,177,178]	Yes [57,179,180]
Camceram®	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.
ChronOS™ Inject	N.D.	N.D.	N.D.	N.D.	Yes [58,62,181]	Yes [58,62,181]
HydroSet™	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.
norian SRS®	(50)	N.D.	N.D.	N.D.	Yes [59–63,95]	Yes [59–62]/no [63]
<i>Calcium sulphate</i>						
Bone Plast®	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.
MIIG® X3	N.D.	N.D.	N.D.	N.D.	Yes [117–119]	Yes [117–119]
OsteoSet®	N.D.	N.D.	N.D.	N.D.	Yes [120–123]/no [124,125]	Yes [120–125,128]
Stimulan®	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.
<i>Bioactive glass</i>						
Cortoss®	(1)	N.D.	N.D.	N.D.	Yes [139]	Yes [140]

Data obtained from the suppliers are given between brackets.
N.D., no data available.

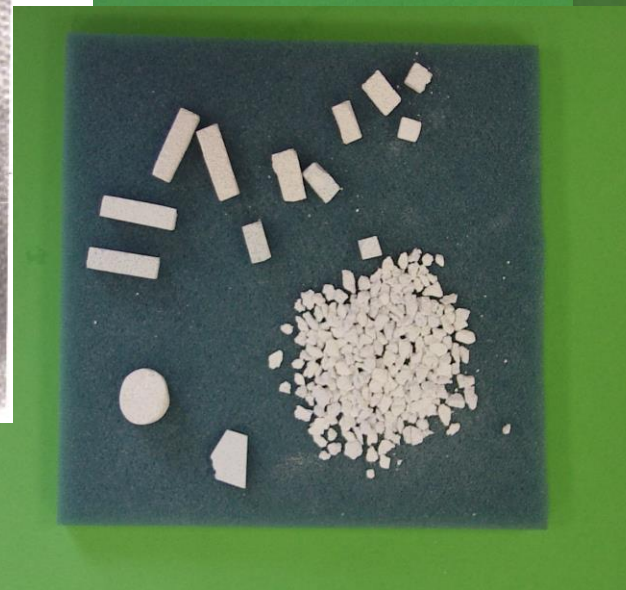
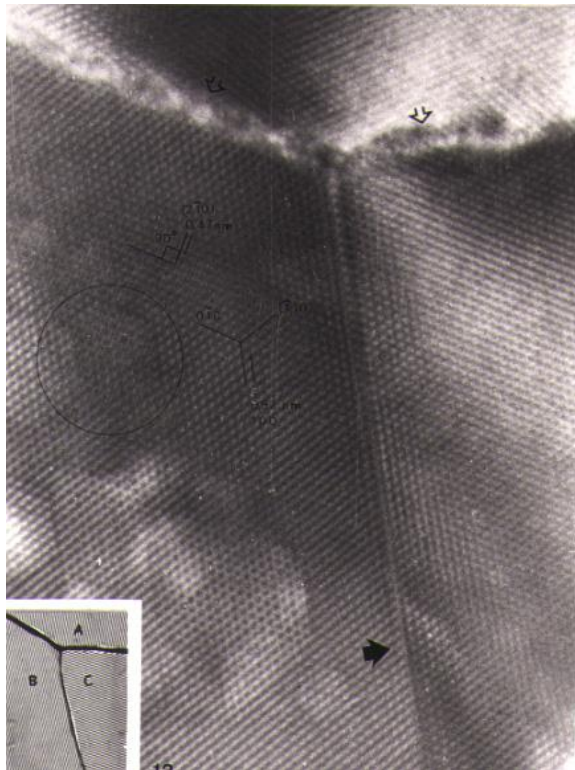
Calcium Phosphate Ceramics

Céramique = cohésion de la structure assurés par traitement thermique



Calcium phosphate ceramics biomaterials

Sinter the powders



Porosity

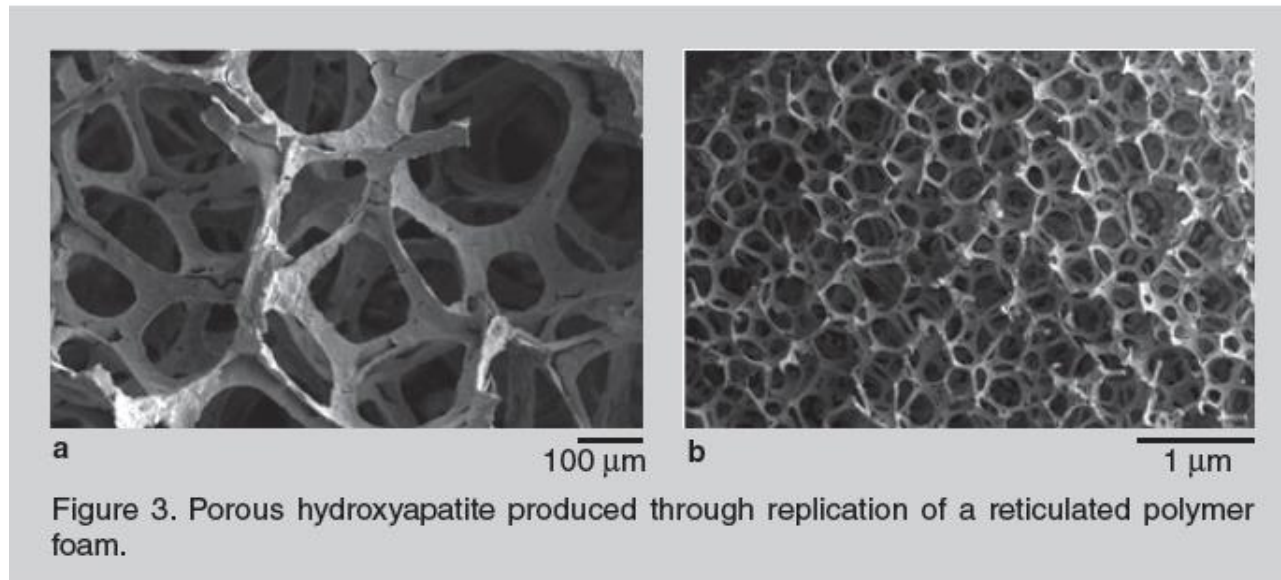
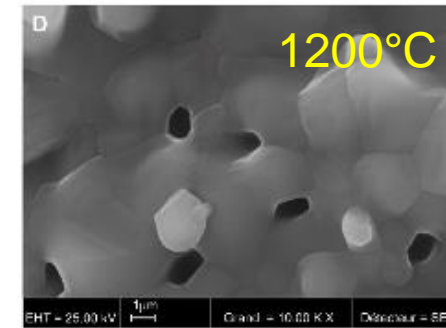
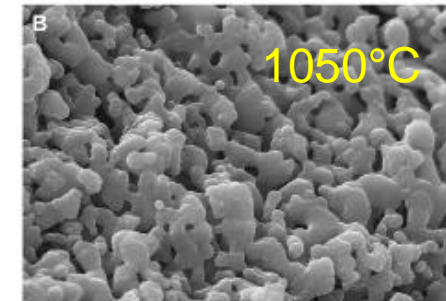
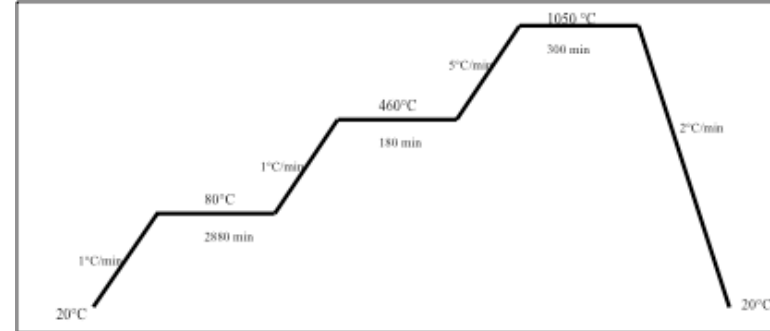
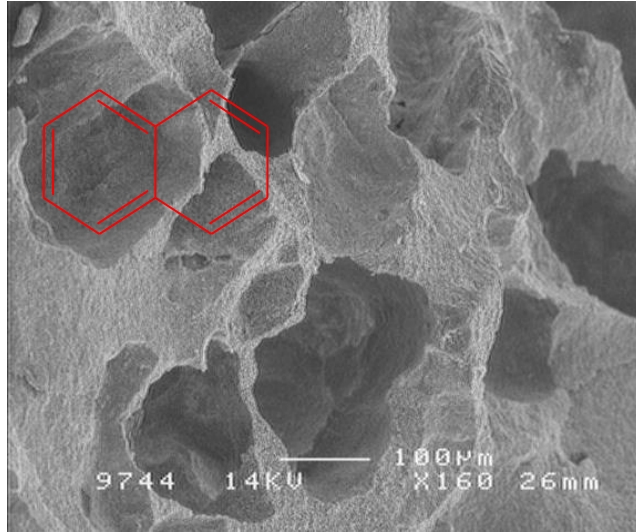


Figure 3. Porous hydroxyapatite produced through replication of a reticulated polymer foam.



Bioglass: 10 milestones from concept to commerce

Larry L. Hench

Department of Biomedical Engineering, Florida Institute of Technology, Melbourne, FL, USA



Bioglass

**Table 1**

Milestones of science and clinical product development of 45S5 Bioglass.

1971	First publication of bonding of bone to bioactive glasses and glass-ceramics
1981	Discovery of soft connective tissue bonding to 45S5 Bioglass
1981	Toxicology and biocompatibility studies (16 in vitro and in vivo) published to establish safety for FDA clearance of bioglass products
1985	First medical product (Bioglass Ossicular Reconstruction Prosthesis) (MEP) cleared by FDA via the 510 (k) process
1987	Discovery of osteostimulation in use of Bioglass particulate in regeneration of bone
1988	Bioglass Endosseous Ridge Maintenance Implant (ERMI) cleared by FDA via the 510 (k) process
1991	Development of sol-gel processing method for making bioactive gel-glasses extending the bioactive compositional range of bioactivity
1993	Bioglass particulate for use in bone grafting to restore bone loss from periodontal disease in infrabony defects (PerioGlas) cleared by FDA via the 510 (k) process
1996	Use of PerioGlas for bone grafts in tooth extraction sites and alveolar ridge augmentation cleared by FDA via the 510 (k) process
2000	FDA clearance for use of NovaBone in general orthopedic bone grafting in non-load bearing sites
2000	Quantitative comparison of rate of trabecular bone formation in presence of Bioglass granules versus synthetic HA and A/W glass-ceramic
2000	Analysis of use of 45S5 Bioglass ionic dissolution products to control osteoblast cell cycles
2001	Gene expression profiling of 45S5 Bioglass ionic dissolution products to enhance osteogenesis
2004	FDA clearance of 45S5 particulate for use in dentinal hypersensitivity treatment (NovaMin)

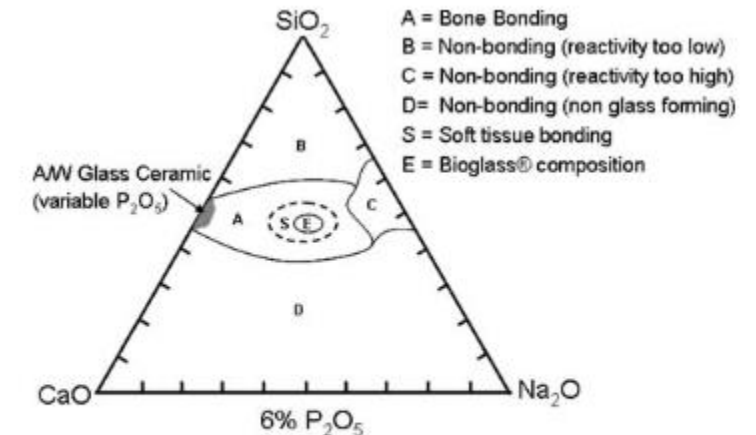


Fig. 1 Compositional diagram for bone-bonding. Note regions A, B, C, D. Region S is a region of Class A bioactivity where bioactive glasses bond to both bone and soft tissues and are gene activating

Bio glass

«A glass is a non-crystalline solid exhibiting the glass transition phenomenon » (Zarzycki 1982)

Selected properties of melt-derived Bioglass 45S5 [60,148].

Property	Value
Density	2.7 g cm ⁻³
Network connectivity	2.12
Glass transition temperature	538 °C
Onset of crystallization	677 °C
Thermal expansion coefficient	15.1 × 10 ⁻⁶ °C ⁻¹
Young's modulus (stiffness)	35 MPa

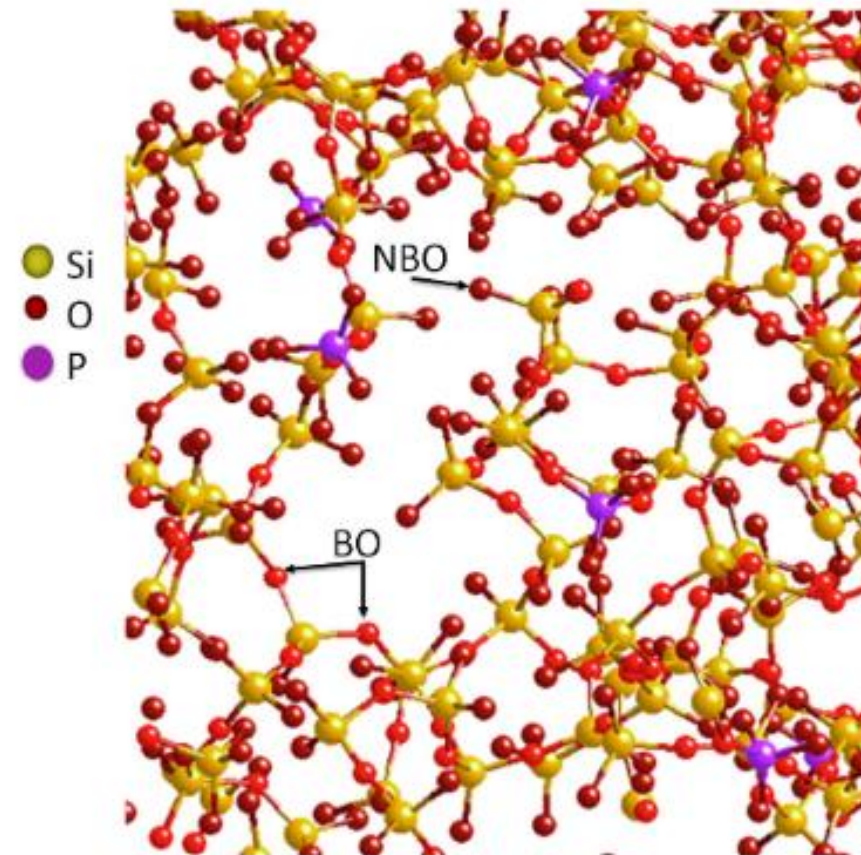


Fig. 10. Section of a model of Bioglass® 45S5, with the Na and Ca ions removed for clarity. NBO – non-bridging oxygen, BO – bridging oxygen. Modified from Cormack et al. [136].

Review

Reprint of: Review of bioactive glass: From Hench to hybrids

Julian R. Jones*

Department of Materials, Imperial College London, South Kensington Campus, London SW7 2AZ, UK



Synthesis

Fusion : Mixture of inorganic raw materials heated between 1300 and 1600 °C then cooled without allowing crystallization

Sol-gel: A solution that gels by polymerizing in an aqueous medium. Then the gel is dried and heated

A flow chart of the acid-catalysed sol-gel process of synthesis of a bioactive glass with schematics of the evolution of the gel and its nanoporosity.

Structural transformations of bio-glasses:

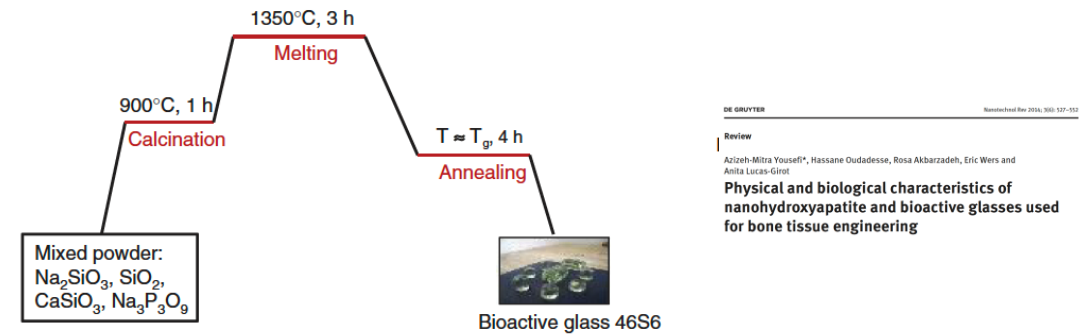
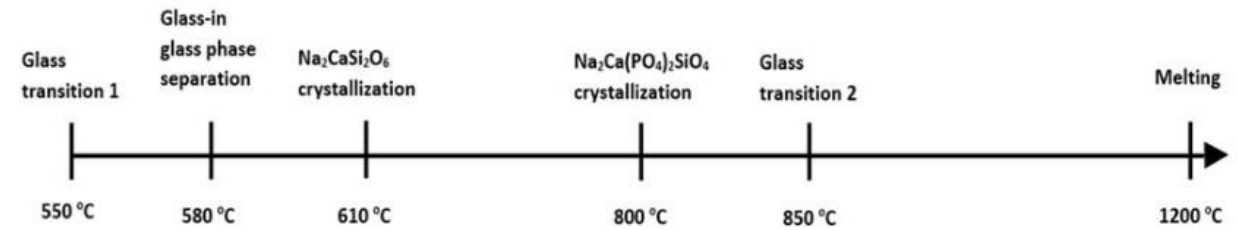
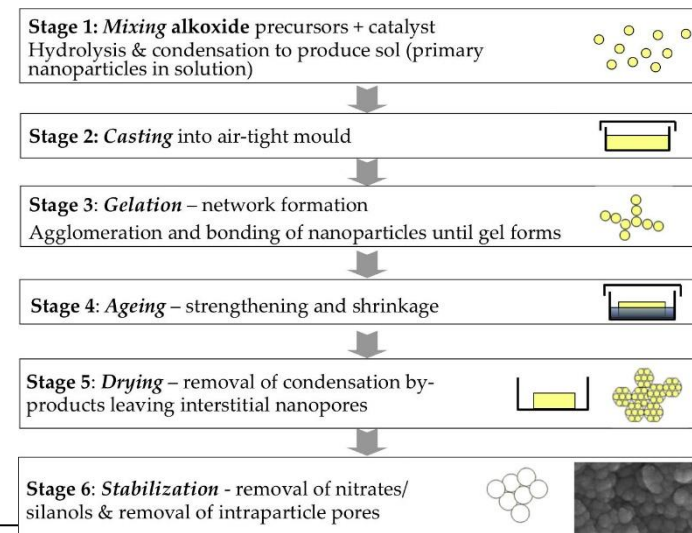


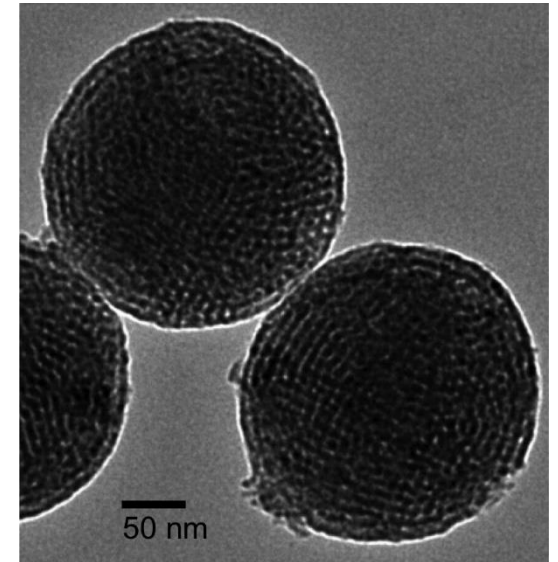
Figure 14 The thermal process for the synthesis of bioactive glass 46S6. Modified from Ref. [138].



DE GRUYTER
Published by De Gruyter
Review
Azizeh-Mitra Yousefi*, Hassane Oudadesse, Rosa Akbarzadeh, Eric Wers and Anita Lucas-Griot
Physical and biological characteristics of nanohydroxyapatite and bioactive glasses used for bone tissue engineering

Different Bio Glasses

- Particle size: Micro to nanometric
- Compositions: with Silver, Potassium, Magnesium, Strontium...
- Nano or meso porosity
- +/- Crystallized



Transmission electron microscope image of ordered mesoporous silica particles. Courtesy of Lijun Ji, Yangzhou University, China.

Table IV. Composition of Bioactive Glasses and Glass-Ceramics (wt%)

Component	45S5 Bioglass®	45S5.4F Bioglass®	45B15S5 Bioglass®	52S4.6 Bioglass®	55S4.3 Bioglass®	KGC Ceravital®	KGS Ceravital®	KGy213 Ceravital®	A/W glass-ceramic	MB glass-ceramic	S45P7
SiO ₂	45	45	30	52	55	46.2	46	38	34.2	19–52	45
P ₂ O ₅	6	6	6	6	6				16.3	4–24	7
CaO	24.5	14.7	24.5	21	19.5	20.2	33	31	44.9	9–3	22
Ca(PO ₃) ₂						25.5	16	13.5			
CaF ₂		9.8							0.5		
MgO						2.9			4.6	5–15	
MgF ₂											
Na ₂ O	24.5	24.5	24.5	21	19.5	4.8	5	4		3–5	24
K ₂ O						0.4				3–5	
Al ₂ O ₃								7		12–33	
B ₂ O ₃			15								2
Ta ₂ O ₅ /TiO ₂								6.5			
Structure	Glass and glass- ceramic	Glass	Glass	Glass		Glass- ceramic	Glass- ceramic	Glass- ceramic	Glass- ceramic	Glass- ceramic	

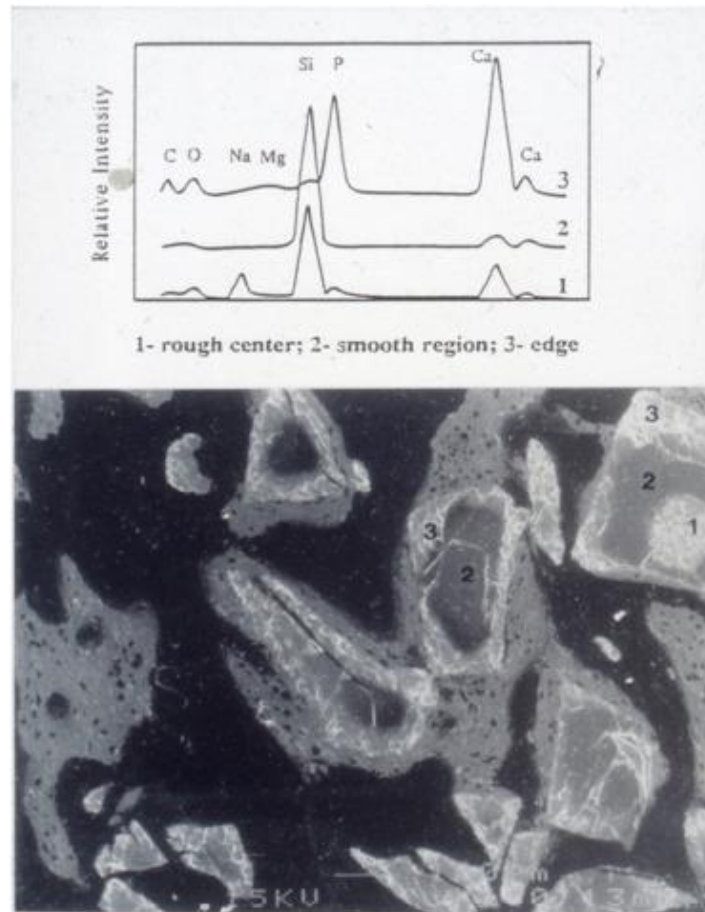


J. Am. Ceram. Soc., 74 [7] 1487–510 (1991)

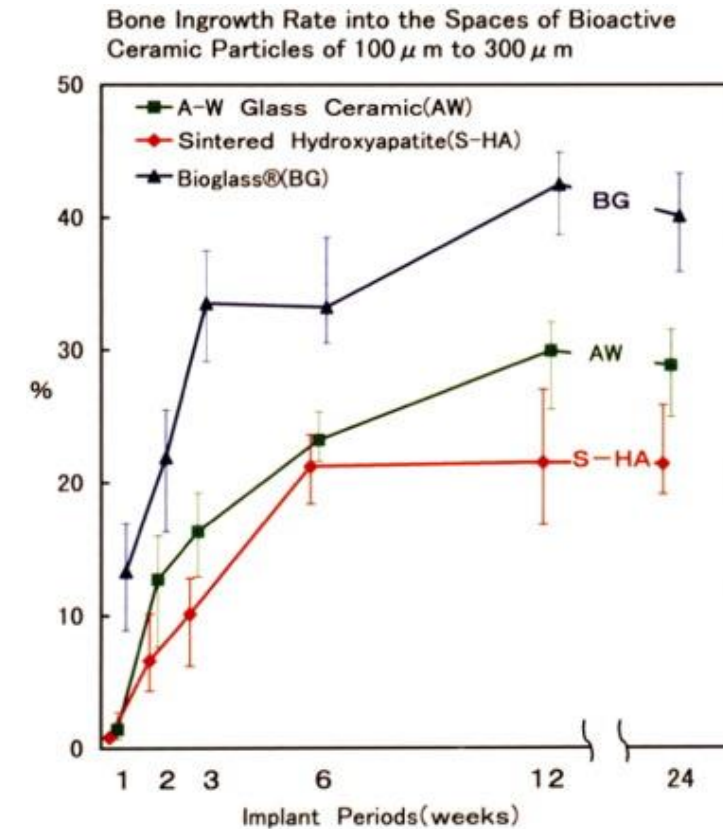
Bioceramics: From Concept to Clinic

Larry L. Hench*

Bone regeneration with Bio Glass

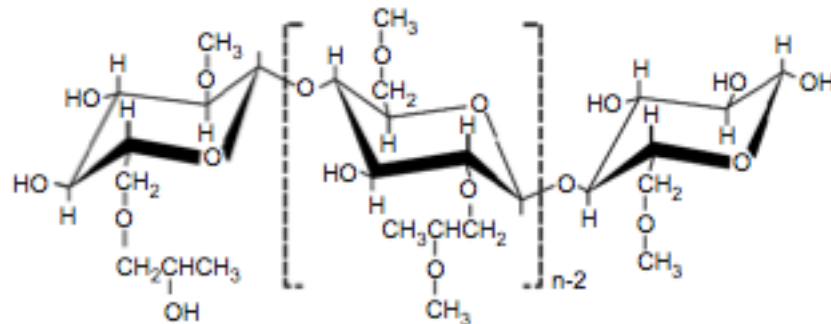


Scanning electron micrograph (SEM) of new bone growing around and connecting 45S5 Bioglass particles to form a regenerated trabecular bone structure. Upper figure is SEM-EDS analysis of the composition gradients of the Bioglass particles due to ion exchange reactions that lead to growth of a biologically active hydroxyl-carbonate-apatite (HCA) layer that is rich in Ca, region #3.



Rapid regeneration of new bone from Class A Bioglass versus Class B bioactive A/W and HA particles in Oonishi critical size defect rabbit condyle model. Note rapid growth of new bone within 4 weeks in the presence of Bioglass particles and eventual greater amount of bone regenerated within the critical size defect.

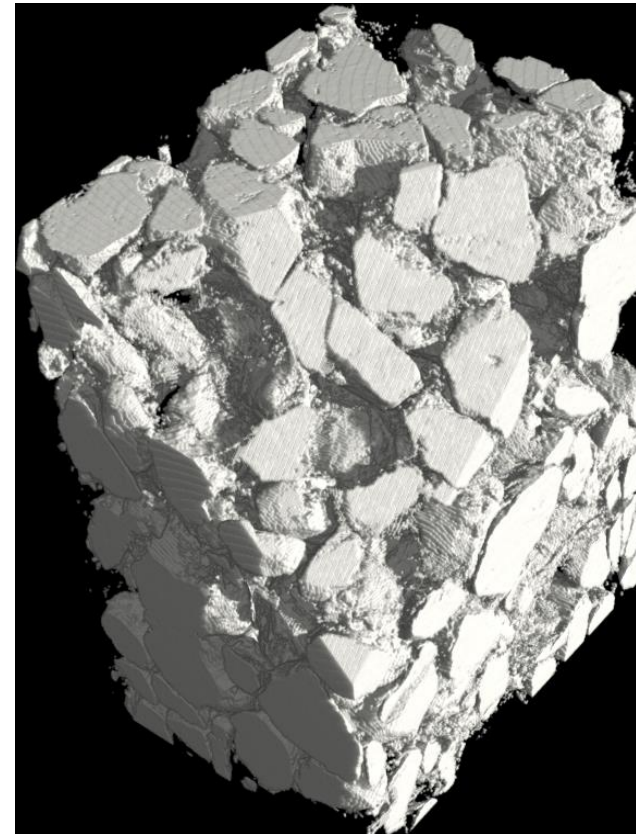
Calcium Phosphate Suspensions in HPMC viscous water solution



2% HPMC in water + BCP granules



WO 9521634 (A1) Injectable Bone Substitute : WEISS P,
DACULSI G., DELECRIN J, GRIMANDI G ET PASSUTI N



Without hardening properties



Available online at www.sciencedirect.com



Biomaterials 28 (2007) 3295–3305

Biomaterials

www.elsevier.com/locate/biomaterials



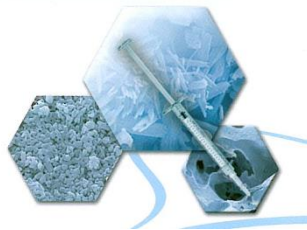
The safety and efficacy of an injectable bone substitute in dental sockets demonstrated in a human clinical trial

Pierre Weiss^{a,b,c,*}, Pierre Layrolle^{a,b}, Léon Philippe Clergeau^{b,c}, Bénédicte Enckel^{b,c},
Paul Pilet^{a,c}, Yves Amouriq^{b,c}, Guy Daculsi^{a,b}, Bernard Giumelli^{b,c}

Pour un acte
simple et efficace

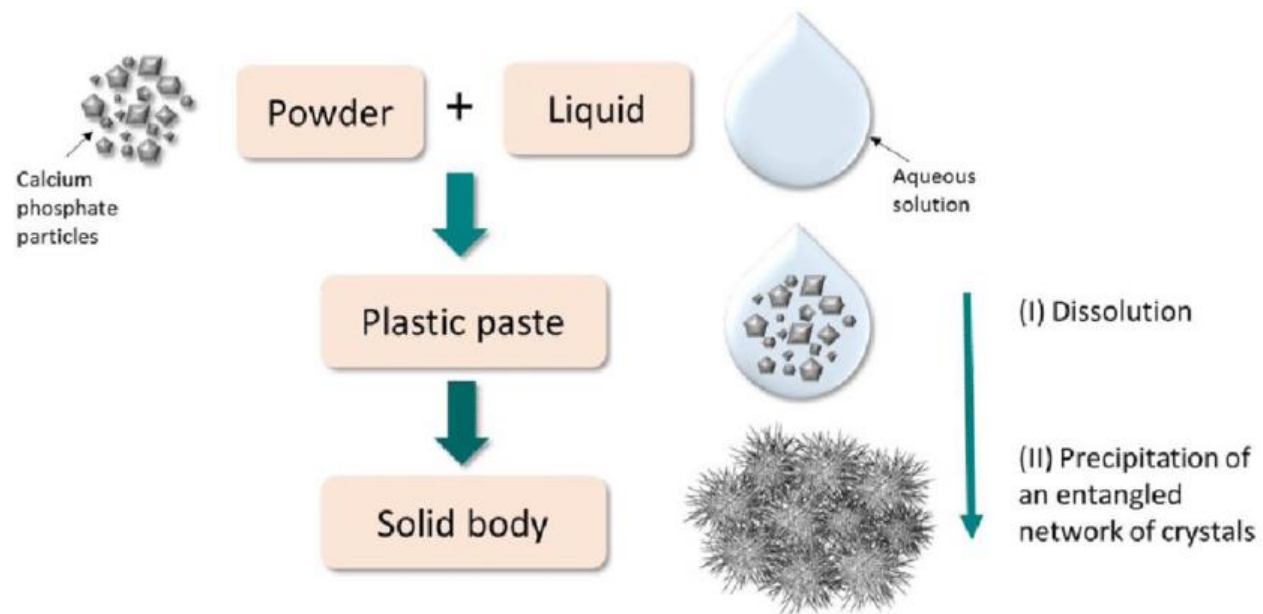
MBCP Gel™

Substitut Osseux Synthétique Injectable

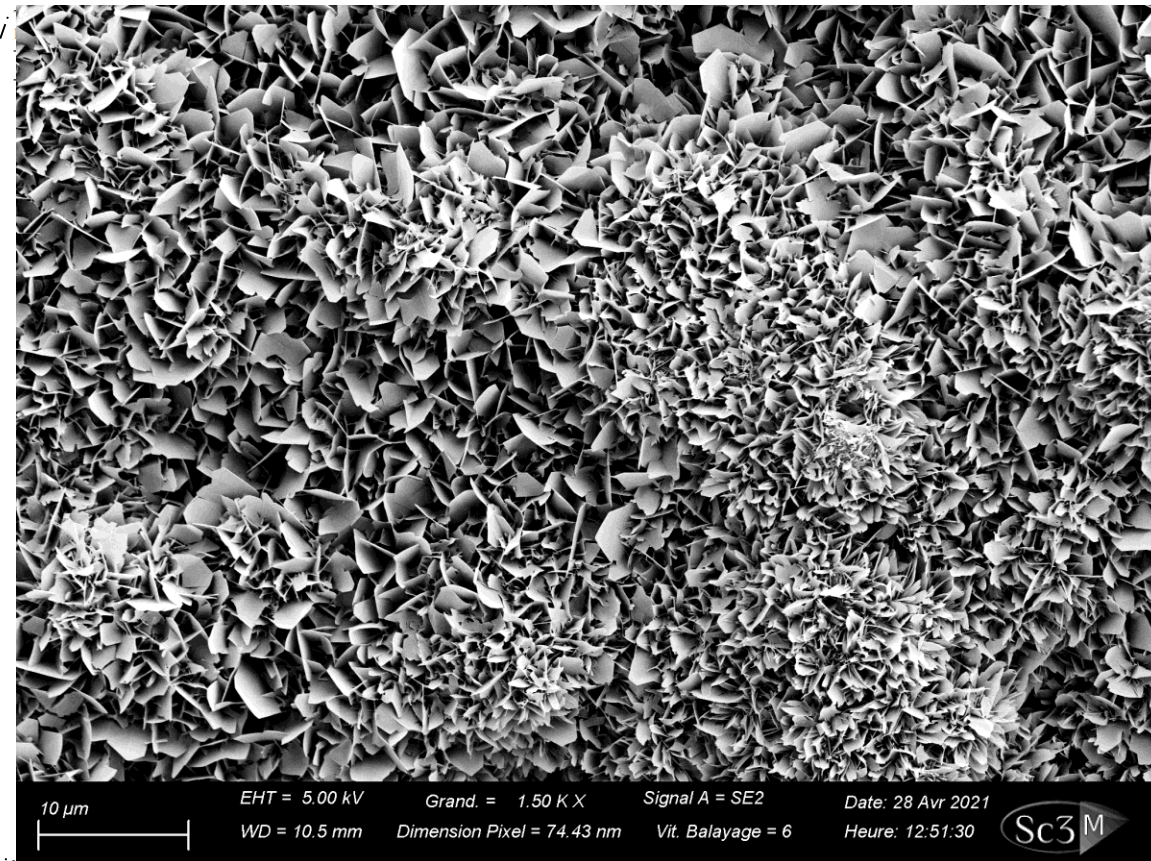


Cement

Cement (hydraulic) = structural cohesion provided by soft chemistry



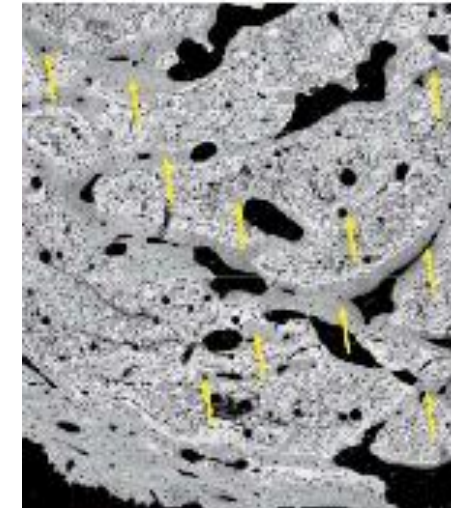
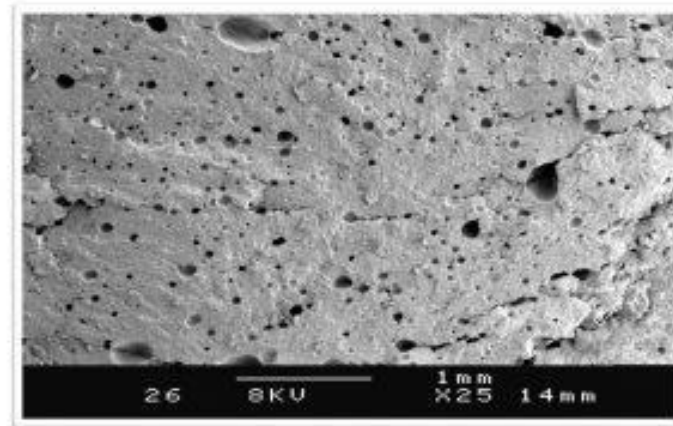
Cements



Macroporous Cement



*Pénétration de Graftys[®]
HBS dans les trabécules*



WO 20081023254 A1, 2008, Macroporous and highly resorbable apatitic calcium-phosphate cement,
WEISS P, KHAIROUN I, BOULER JM

Mixture of a powder :

- 78 wt % α -tricalcium phosphate (α -TCP),
- 5 wt % dicalcium phosphate dihydrate ($\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$)
- 5 wt % monocalcium phosphate mono-hydrate (MCPM)
- 10 wt % calcium-deficient hydroxyapatite (CDA)
- **2 wt % HPMC.**

liquid phase : 5 wt % Na_2HPO_4 aqueous solution

Graftys[®] QuickSet

About Graftys[®] QuickSet

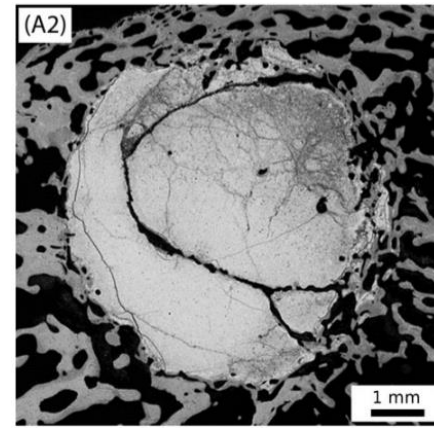
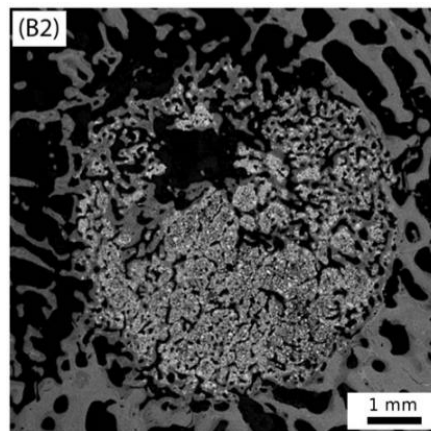
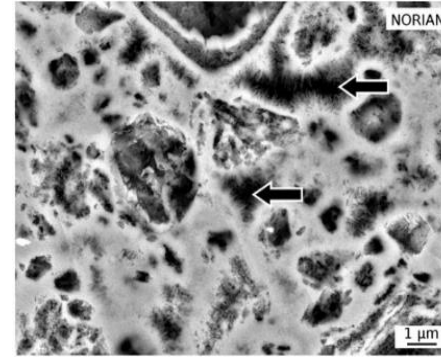
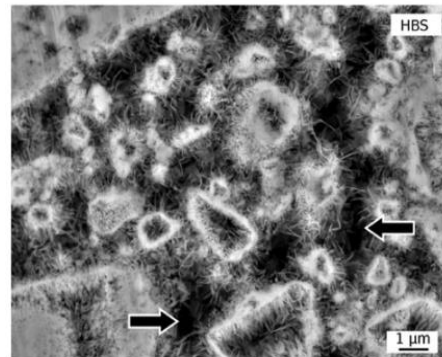
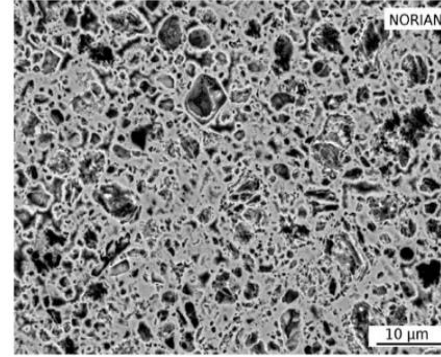
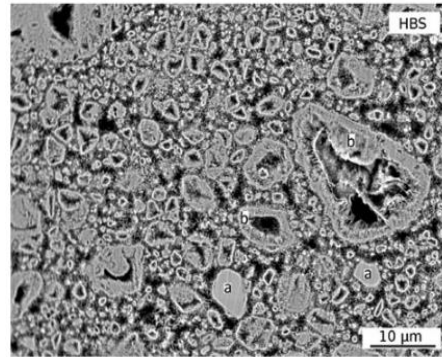
Graftys[®] QuickSet is a Calcium Phosphate Cement of high viscosity with mechanical properties similar to those of cancellous bone, supplied in a user-friendly double-syringe, which is pre-filled with a powder (calcium phosphate salts and HydroxyPropylMethylCellulose (HPMC)) and with a phosphate-based (Na_2HPO_4) aqueous solution. Mixing of these two components in the syringe results in an injectable paste, which hardens under *in vivo*. The final product formed during the setting reaction is a calcium-deficient apatite CDA that closely mimics the mineral phase of natural bone.



Total porosity measured by Hg intrusion (%)

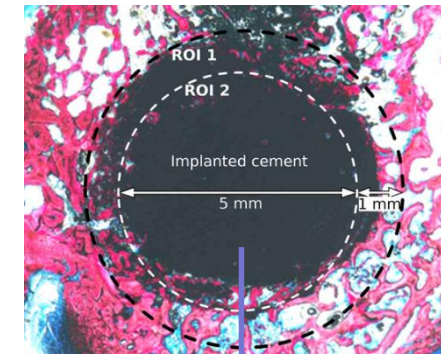
61 ± 6 /

44 ± 4



In vivo resorption of injectable apatitic calcium phosphate cements: Critical role of the intergranular microstructure

Myriam Le Ferrec^{1,2} | Charlotte Mellier¹ | Francois-Xavier Lefèvre³ |
Florian Boukhechba¹ | Pascal Janvier³ | Gilles Montavon² | Jean-Michel Bouler³ |
Olivier Gauthier^{4,5} | Bruno Bujoli³



Implantation time	HBS		NORIAN	
	Cement area density (%)	Bone area density (%)	Cement area density (%)	Bone area density (%)
4 weeks	88.6 ± 13.2	5.6 ± 1.7*	99.0 ± 0.9	0.6 ± 0.2
8 weeks	76.9 ± 15.5*	13.5 ± 4.1*	98.9 ± 0.9	0.8 ± 0.2
12 weeks	79.1 ± 15.7*	11.0 ± 3.3*	97.5 ± 0.9	1.5 ± 0.5

8 weeks of implantation
in Rabbit bone

Commercialized injectable bone substitutes

Biological properties

Mechanical properties

In'Oss™ MBCP™



MBCP™ Putty - In'Oss™

Hydroxyapatite + Phosphate
Tricalcique Bêta (βTCP)
+ viscous liquid = 2% HPMC



WO 9521634 (A1) Injectable Bone Substitute : WEISS P, DACULSI G., DELECRIN J, GRIMANDI G ET PASSUTI N

Injectable suspensions

- + Biocompatibility
- + Injectability
- + Osteoconduction
- Mechanical properties

Solid materials

Graftys® QuickSet



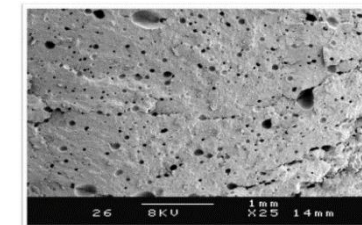
Transfer

Mix : 2 min

Inject

- + Biocompatibility
- + Injectability
- + mechanical properties

- Porosity = HPMC particles
- Osteoconduction



calcium phosphate salts +
HydroxyPropylMethylCellulose (HPMC) + Na₂HPO₄



WO 20081023254 A1, 2008, Macroporous and highly resorbable apatitic calcium-phosphate cement, WEISS P, KHAIROUN I, BOULER JM

Injectable macromolecule-based calcium phosphate bone substitutes

Hilal Moussi,^{1b} Pierre Weiss,^{1b,*} Jean Le Bideau,^{1b} H el ene Gautier^{1b} and Baptiste Charbonnier¹

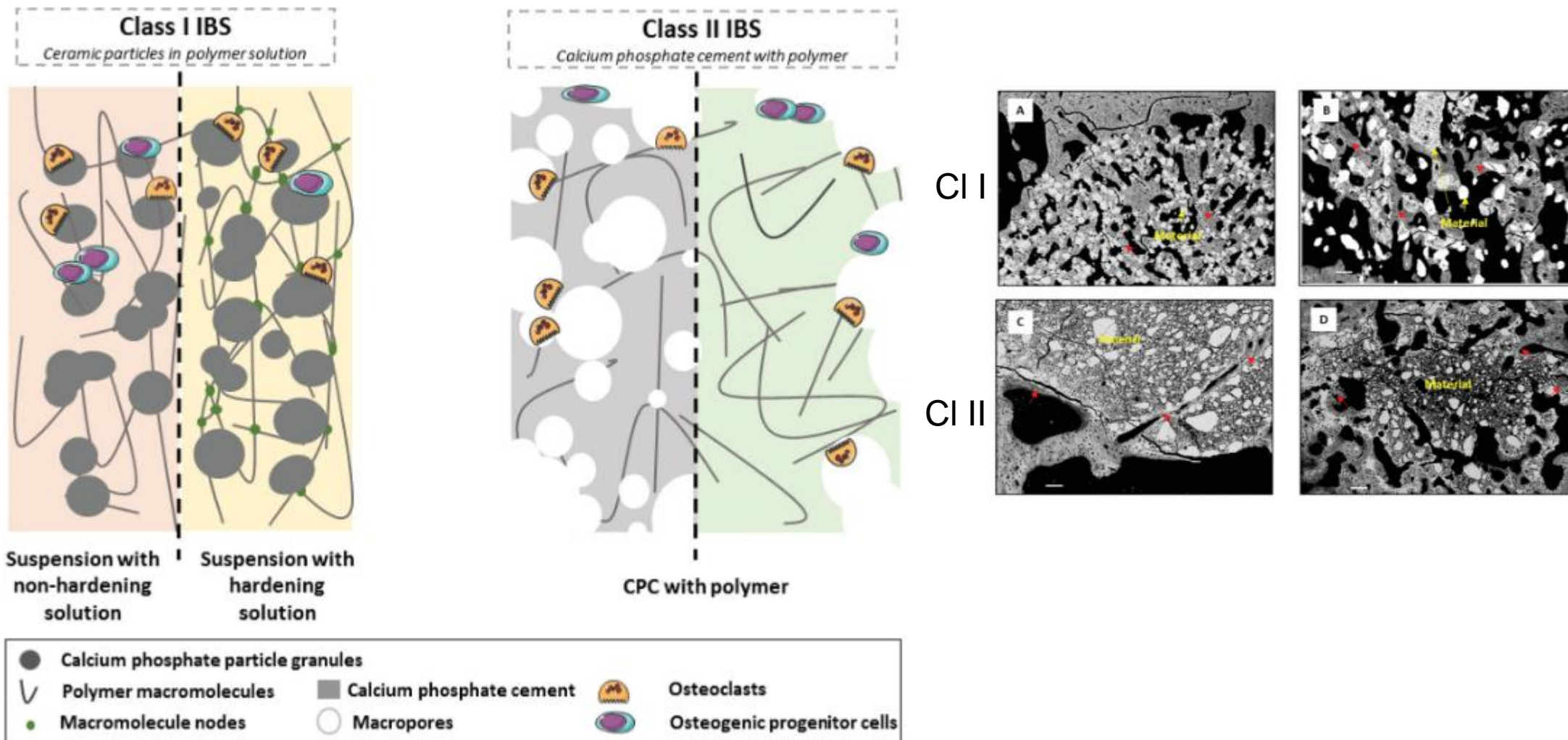


Fig. 1 The two classes of IBS present different osteoconduction mechanisms: for Class I, calcium phosphate supports are made of divided ceramics (thermally-sintered particles, resulting in high density materials); for Class II, calcium phosphate supports are made of bulk cements (soft chemistry route to interlinked particles, resulting in low-density materials showing macropores).



Injectable macromolecule-based calcium phosphate bone substitutes

Hilel Moussi,^{ab} Pierre Weiss,^{ic*} Jean Le Bideau,^{id} Hélène Gautier^{id} and Baptiste Charbonnier^a

Cite this: *Mater. Adv.*, 2022, 3, 6125

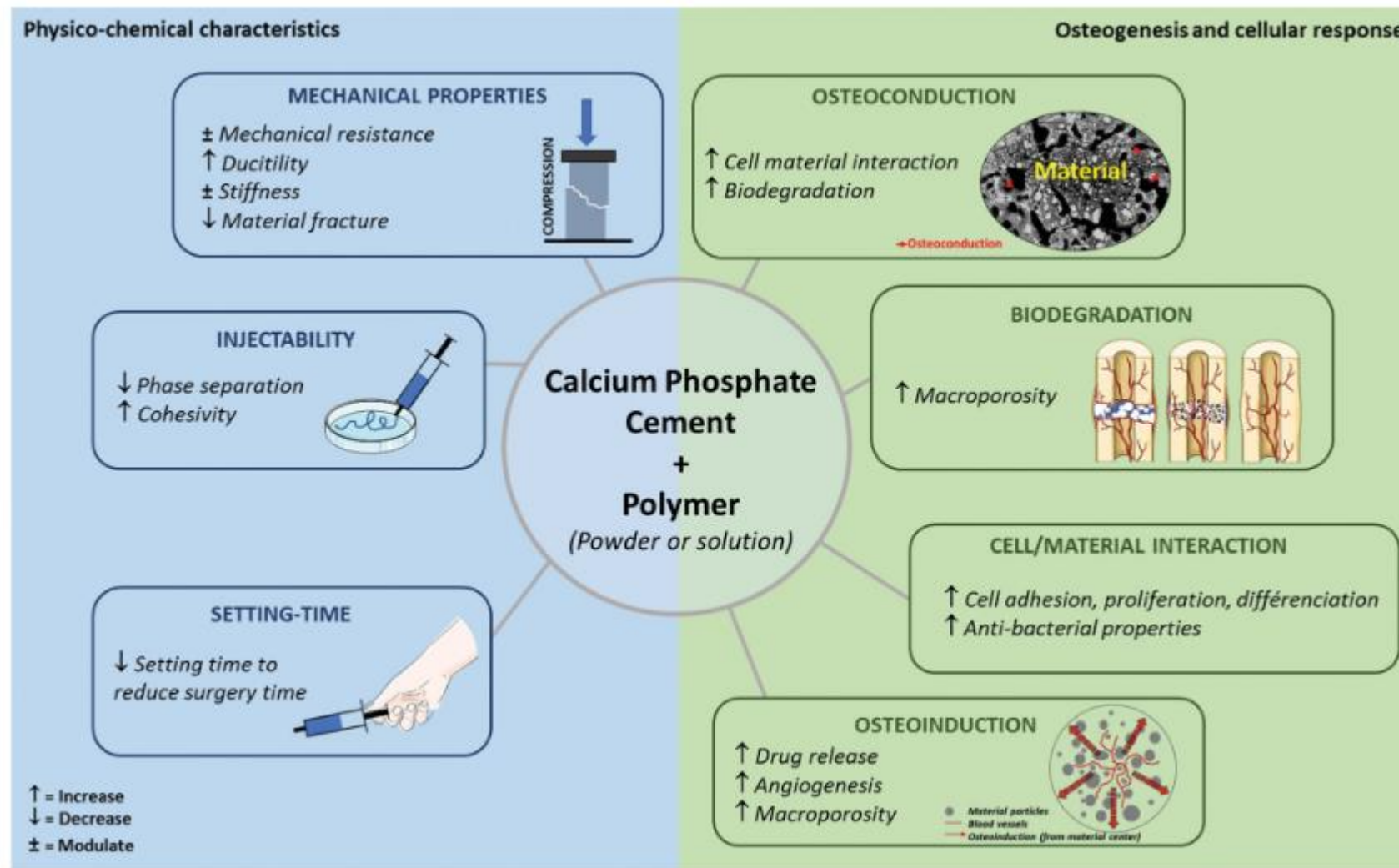
Table 1 Class I and Class II injectable and extrudable commercialized materials currently on the market (TCP = tricalcium phosphate, HA = hydroxyapatite, CaP = calcium phosphate, CDHA = calcium deficient hydroxyapatite, HPMC = hydropropylmethylcellulose, rhBMP = recombinant human bone morphogenetic protein)

Commercial name and manufacturer	Material class	Inorganic phase	Organic phase	Application	Characteristics	Clinical trial number and reference
MBCP-Gel/In'Oss™ (Biomatlante)	I	CaP granules	HPMC polymer solution	Filling osseous defects of various origins	Ready to use, cohesivity/putty	NCT00740311; (ref. 30)
ExcelOs-inject (CGBIO)	I	Beta TCP granules	Biodegradable hydrogel with or without rhBMP-2	Spinal fusion	Ready to use, injectable, cohesivity	NCT02714829; (ref. 31)
Mastergraft (Medtronic)	I	β-TCP + HA	Bovine collagen matrix	Spinal fusion	Putty	NCT01491542; (ref. 32)
Vitoss (Stryker)	I	β-TCP granules	Bone marrow aspirate	Spinal pathologies	Putty ultra-porous, flexible	NCT03509480; (ref. 33)
ChronOS (DePuy Synthes)	I	β-TCP granules	Autologous blood and/or bone marrow	Tibial plateau fractures	Putty	NCT02056834; (ref. 34)
ChronOS Inject (DePuy Synthes)	I	Brushite matrix and (TCP) granules		Proximal tibial fractures	Injectable, degradable	NCT02056834; (ref. 35)
CERAMENT® (Bone Void Filler)	I	HA and calcium sulfate cement		Tibial plateau fractures	Injectable, degradable	NCT01828905; (ref. 36)
Norian® Drillable (DePuy Synthes)	II	Carbonated apatite cement	Bioresorbable polylactide/glycolide copolymer fibres	Tibial fractures	Mechanical resistance 24 hours after injection	NCT01132508; (ref. 37)
Graftys quickset/HBS (Graftys)	II	CDHA cement	Powder of HPMC polymer	Bone disease, bone fractures	Porosity	NCT02575352; (ref. 38)

Injectable macromolecule-based calcium phosphate bone substitutes

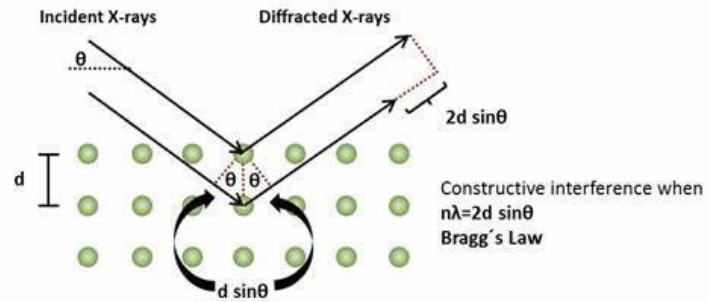
Hilal Moussi,^{ab} Pierre Weiss,^{b*} Jean Le Bideau,^b Hélène Gautier^b and Baptiste Charbonnier^a

Cite this: *Mater. Adv.*, 2022, 3, 6125

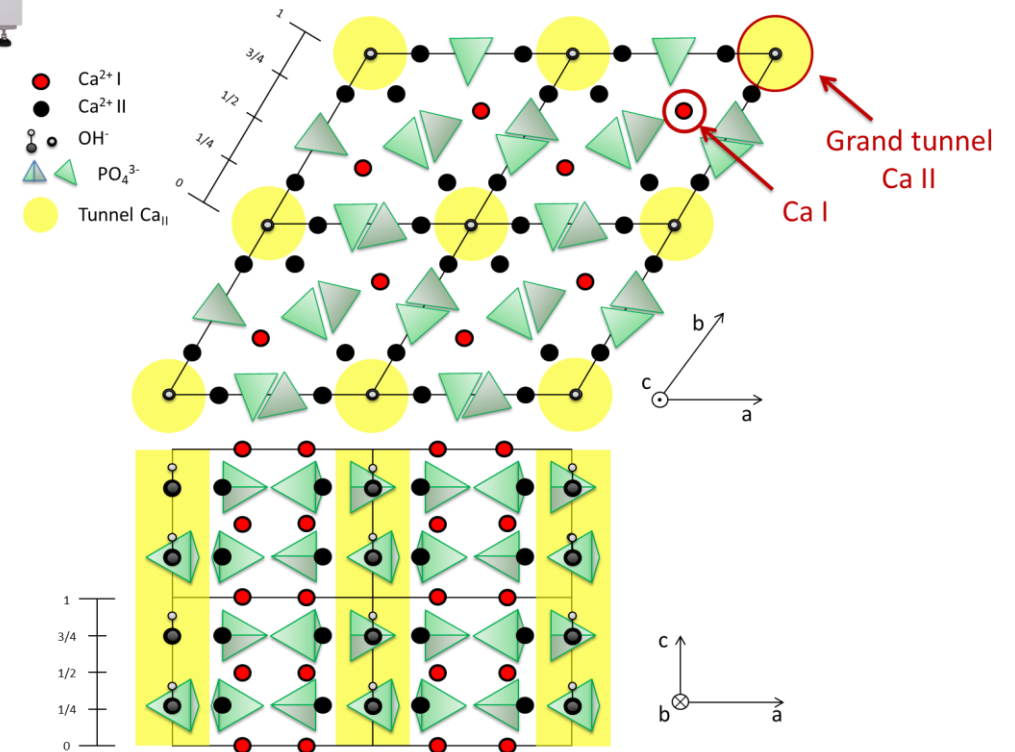
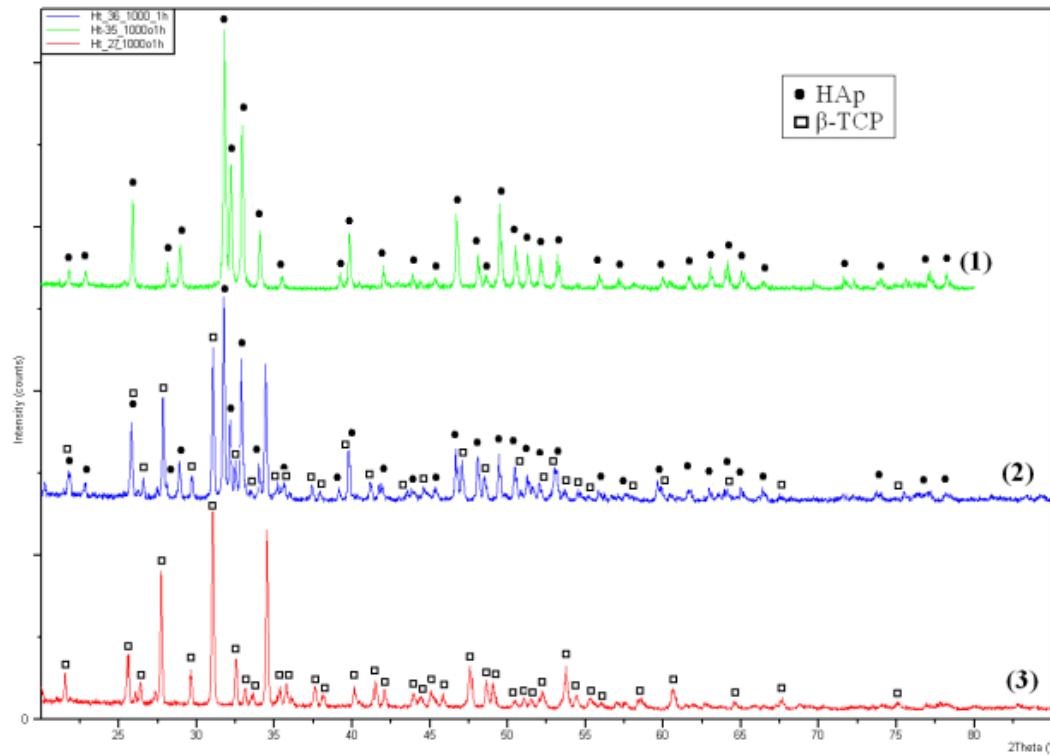


Modulation of IBS properties by a combination of polymers with calcium phosphate cement (Class II).

Physico-chemical characterization



X-ray diffraction



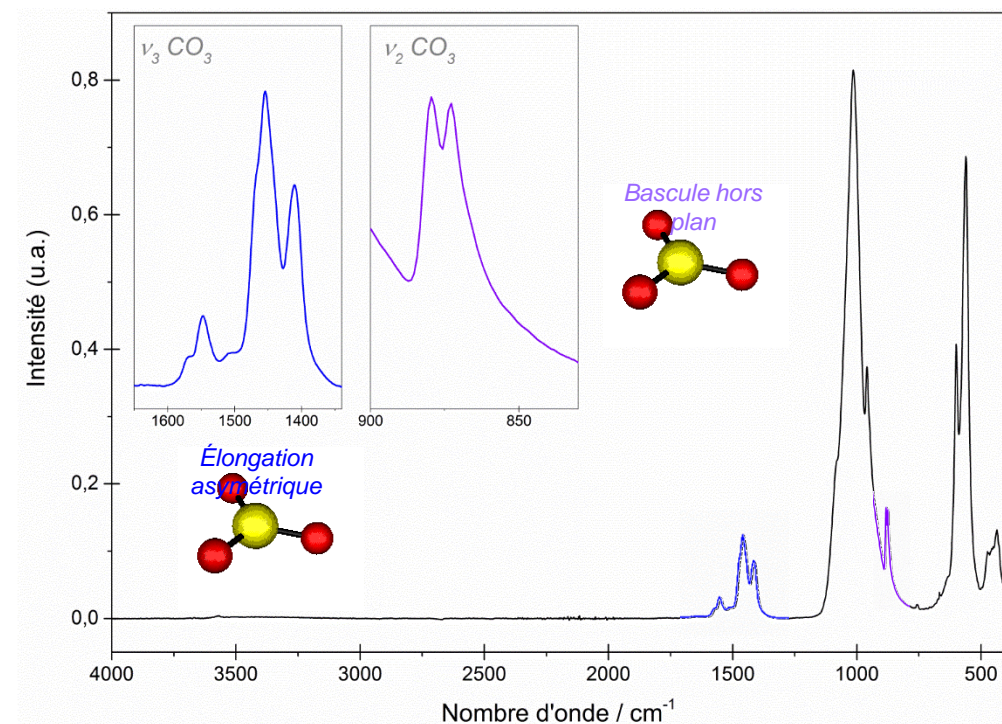
Physico-chemical characterization

Infrared Spectroscopie



Bandes IR (cm-1)	Intensité	Attribution	
3700 - 3000	moyenne	H-O-H, H2O adsorbée	
3640 et 3540	faible	O-H, des groupes Ca(OH)2	
3580	épaulement	O-H, vibration de valence symétrique des ions OH ⁻	v ₅
1630	moyenne	H2O et CO2 adsorbés	
1400 - 1380 *	forte	N-O des groupes NO3	
1500 - 1350	faible	C-O des groupes CO3	v ₃
1200 - 1180	épaulement	P-OH, groupes HPO4	
1120 - 1020	forte	P-O, élongation antisymétrique des ions PO4 ³⁻	v ₃
965 - 960	moyenne	P-O, élongation symétrique des ions PO4 ³⁻	v ₁
900 - 850	faible	C-O des groupes CO3	v ₂
880 - 865	faible	Elongation P-OH des groupes HPO4	
820 *	faible	N-O des groupes NO3	
630	épaulement	O-H mouvement de libration des ions OH ⁻	v _L
600 - 550	forte	P-O, déformation antisymétrique des ions PO4 ³⁻	v ₄
460	faible	P-O, déformation symétrique des ions PO4 ³⁻	v ₂

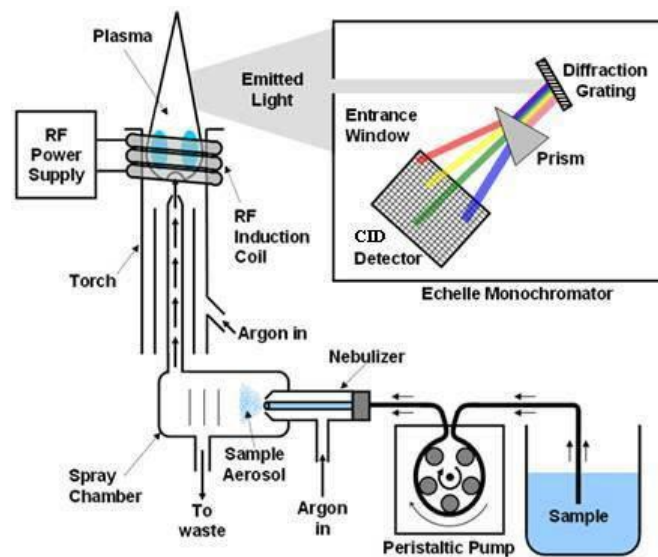
*Résidus nitrés de synthèse présents dans une poudre immédiatement après synthèse ; la majorité des nitrates étant éliminés par traitement thermique de la poudre à faible température (T ≈ 400 °C).



Physico-chemical characterization

Analyses élémentaires

ICP-AES (ICP-MS également possible!)

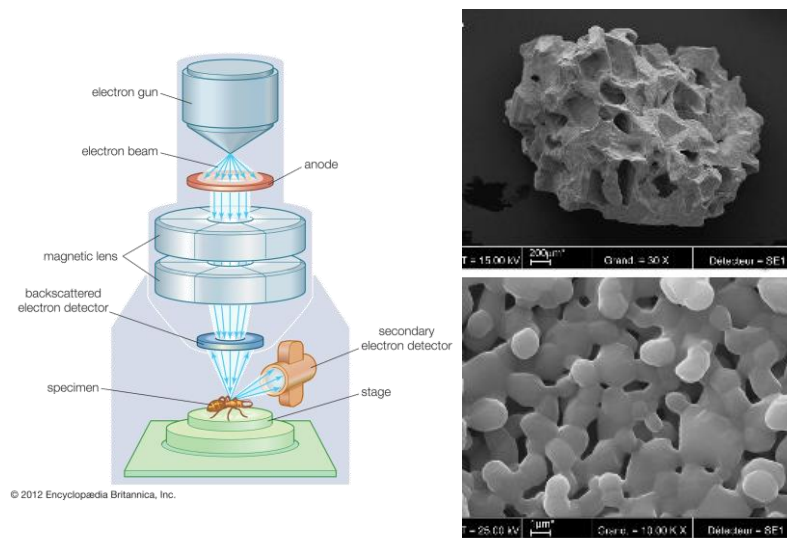


Dosages chimiques (chromato ionique, dosage C,...)

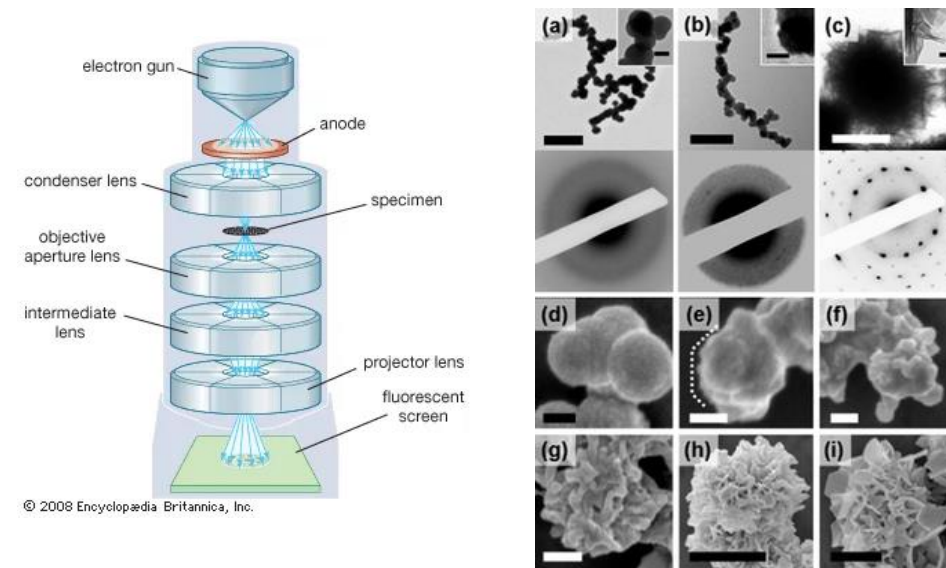


Physico-chemical characterization

Microscopie électronique à balayage



Microscopie électronique en transmission

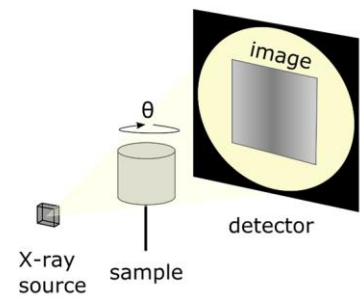


Pan et al., Mystery of the transformation from amorphous calcium phosphate to hydroxyapatite, Chem. Comm. (2010)

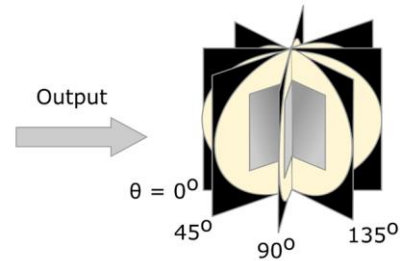
Physico-chemical characterization

Micro/Nano tomographie à rayons X

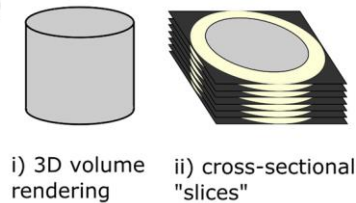
A Image acquisition



B 2D projections of sample

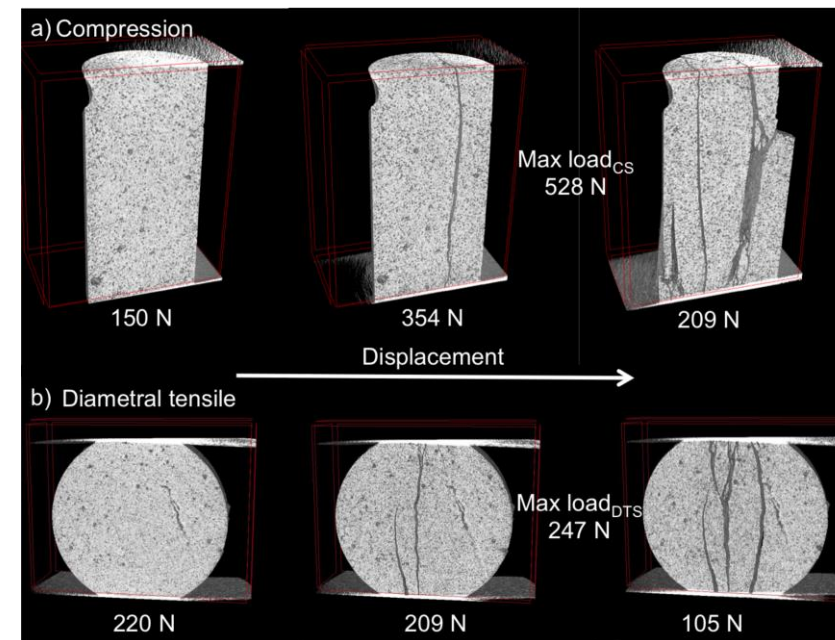


D



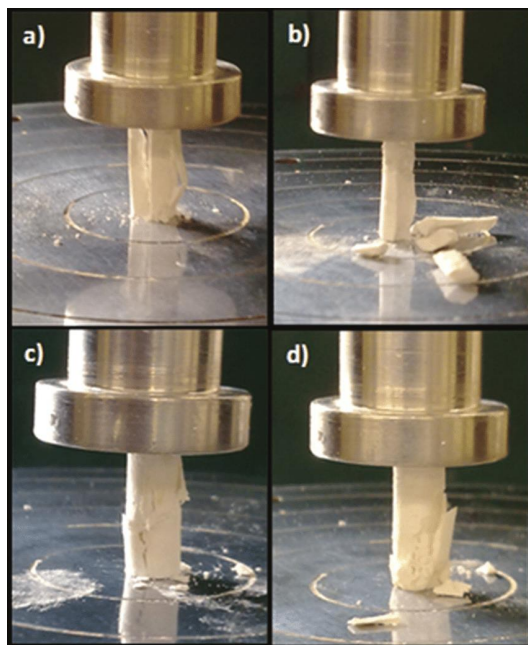
C Reconstruction

- 1) Filtered backprojection
or
- 2) Iterative reconstruction



Physico-chemical characterization

Essais Mécaniques

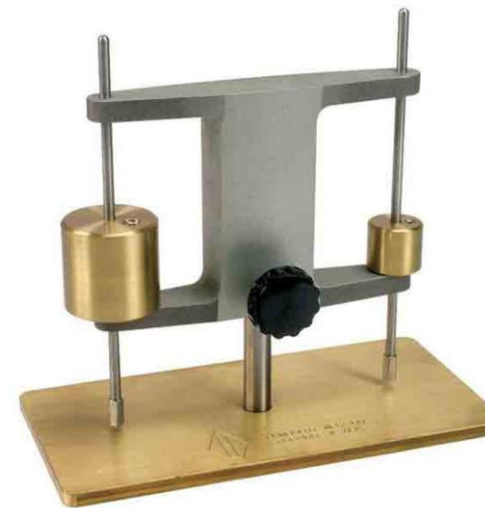
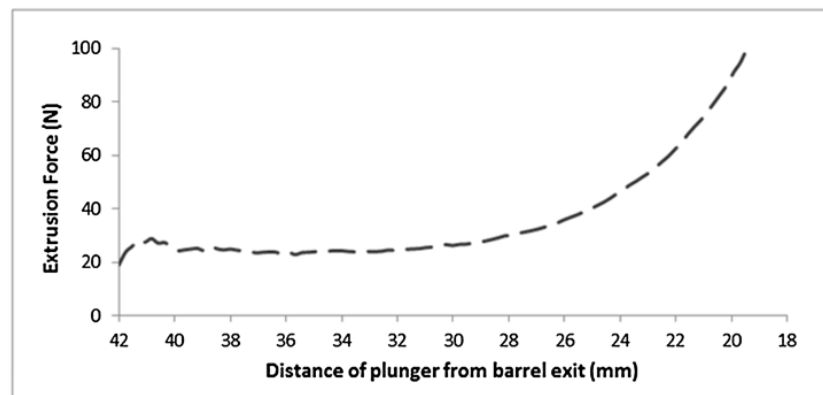
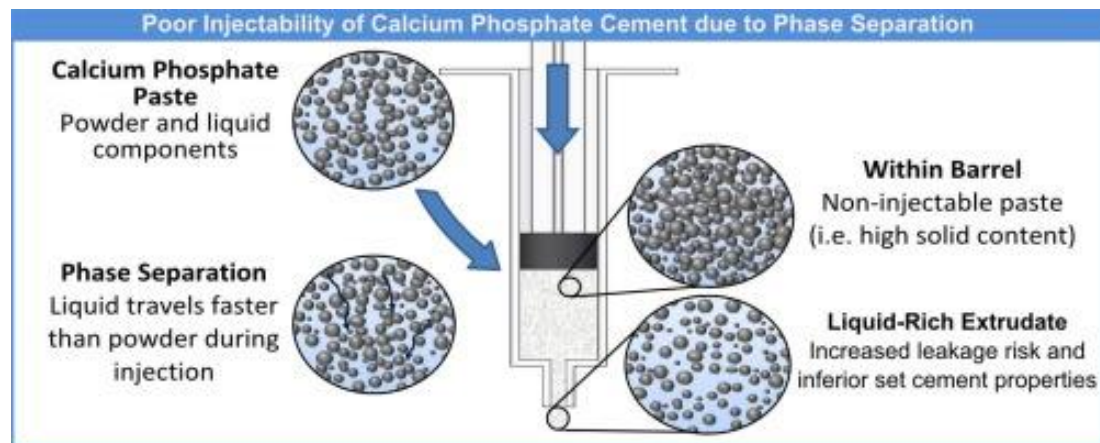


Property	Cortical bone	Calcium phosphate cement
Specific Gravity (g/cm ³)	1,7-2,1	1,7-2,0
Tensile Strength (MPa)	60-160	2,1-14
Compressive strength (MPa)	13-18	20-91
Young's Modulus (GPa)	3-30	35-105
Fracture energy (J/m ²)	390-560	–
Resistance to fracture (Mpa/m ²)	2-12	0,3-0,8
Composition	Inorganic + organic	Inorganic

Source: Adapted from Wagh (2016)

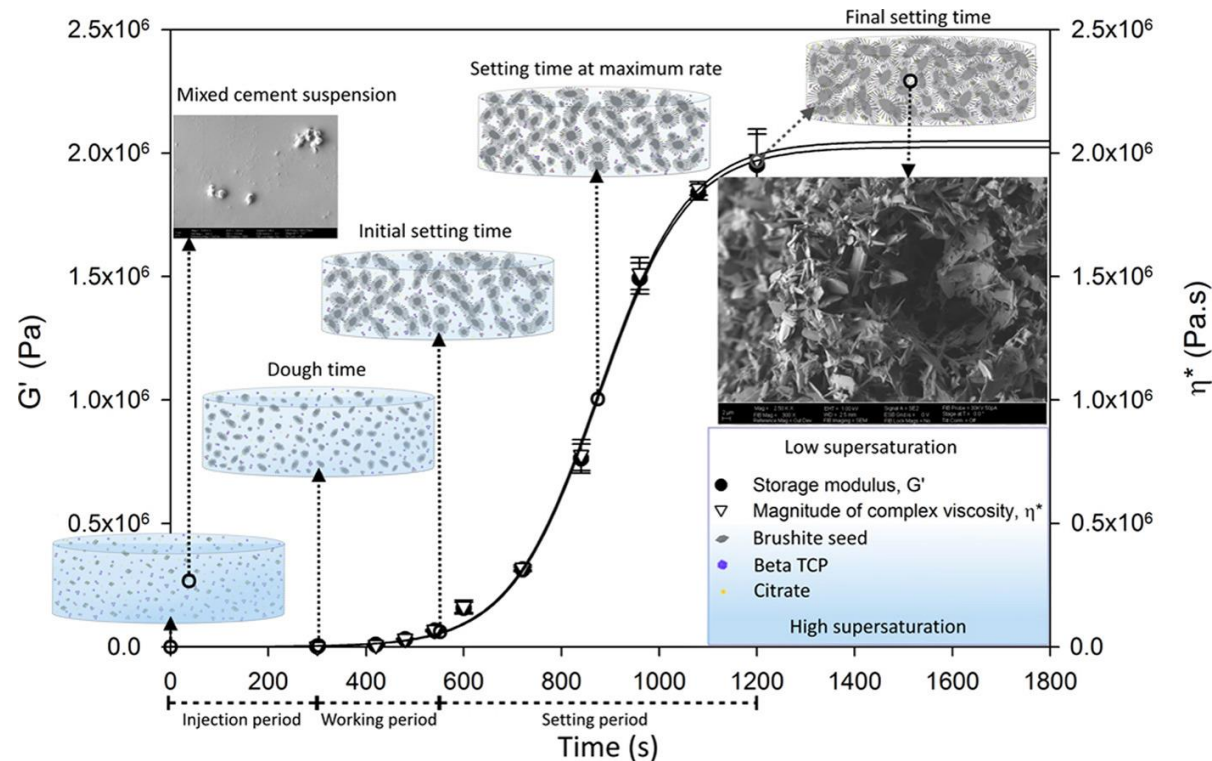
Physico-chemical characterization

Injectabilité et temps de prise (ciment)



Physico-chemical characterization

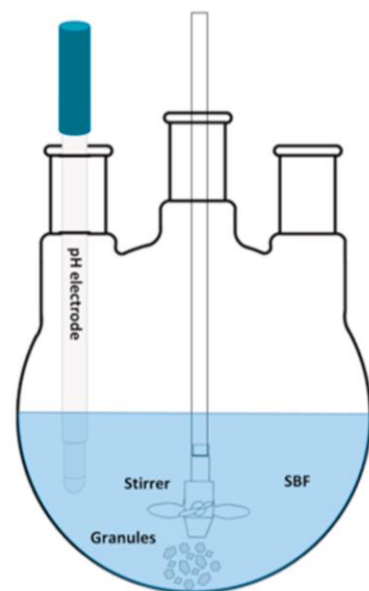
Comportement rhéologique (ciment)



Erdem et al., The rheological behavior of a fast-setting calcium phosphate bone cement and its dependence on deformation conditions (2017)

Physico-chemical characterization

Comportement en milieu biologique simulé



Analyses multimodales

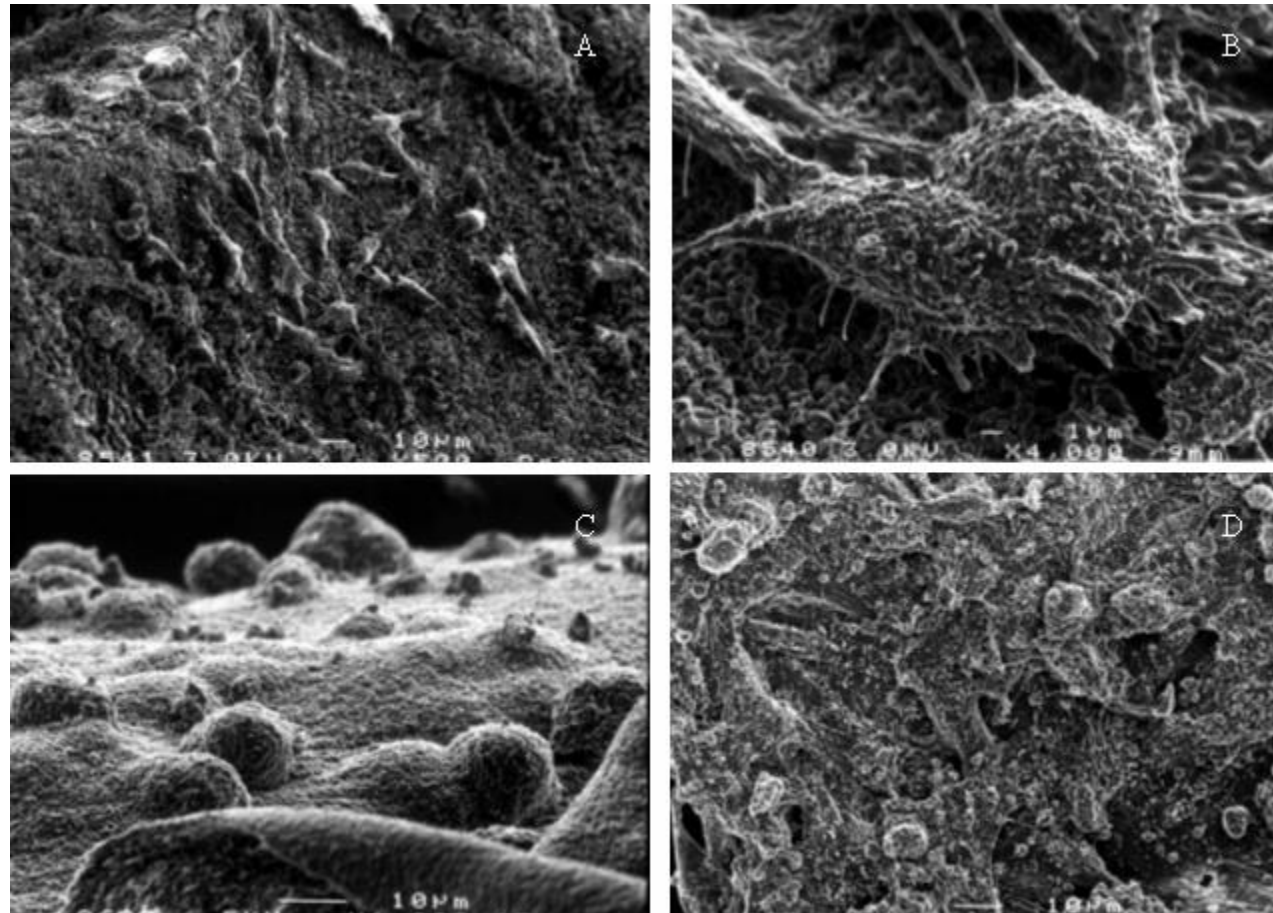


Changement:

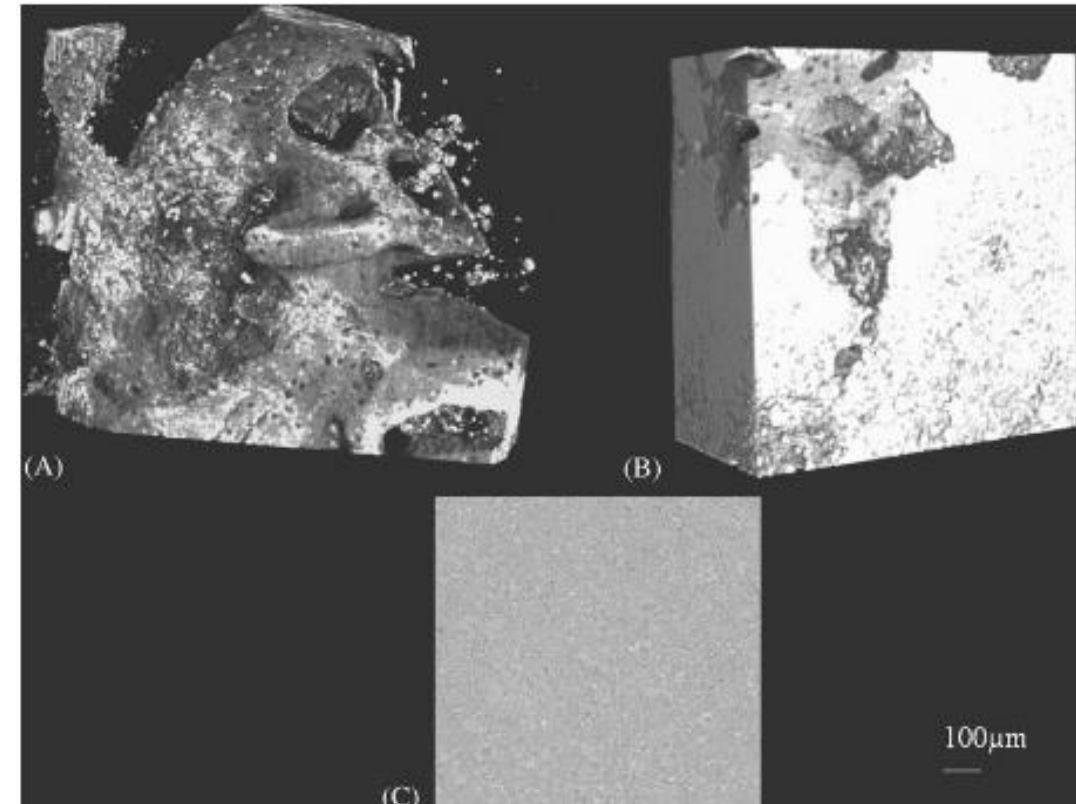
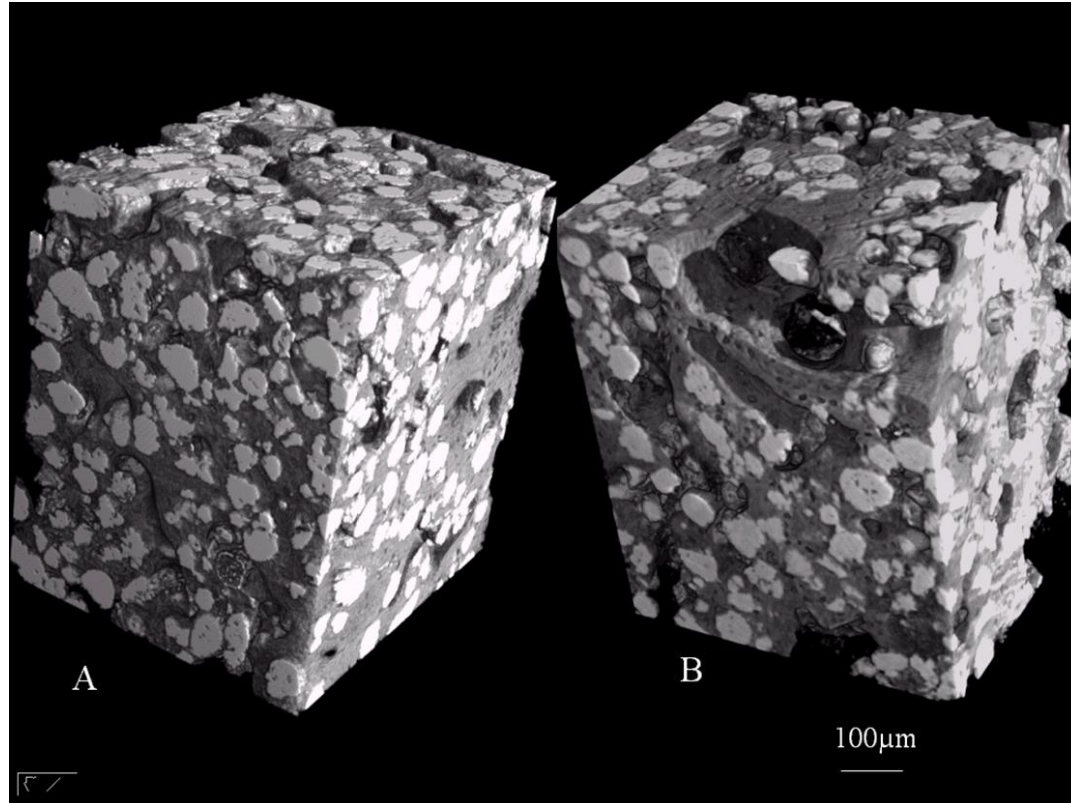
- Composition
- Phases
- Masse
- Porosité
- Surface spécifique
- Morphologie...

Mazzoud et al., Cell-free, quantitative mineralization measurements as a proxy to identify osteoinductive bone graft substitutes, Biomaterials 2022

Scaffold for cells = osteoconduction



Osteoconduction



Available online at www.sciencedirect.com

SCIENCE @ DIRECT®

Biomaterials

Biomaterials 24 (2003) 4591–4601

www.elsevier.com/locate/biomaterials



Synchrotron X-ray microtomography (on a micron scale) provides three-dimensional imaging representation of bone ingrowth in calcium phosphate biomaterials

P. Weiss^{a,*}, L. Obadia^a, D. Magne^a, X. Bourges^a, C. Rau^b, T. Weitkamp^b, I. Khairoun^a, J.M. Bouler^a, D. Chappard^c, O. Gauthier^{a,d}, G. Daculsi^a

Osteoinduction of CaP

Table 1 Histomorphometry of bone formed in porous hydroxyapatite specimens

Animal model	% bone volume	
Rabbit	0.5 ± 0.3	(16)
Dog	0.75 ± 0.49	(8)
Baboon	11.3 ± 3.02	(16)

Bone volume was calculated using a Zeiss Integration Platte II with 100 lattice points superimposed over the centre of the sections, as described in Materials and Methods. The number of specimens per animal species is given in parentheses. Values (in %) are means ± s.e.m.

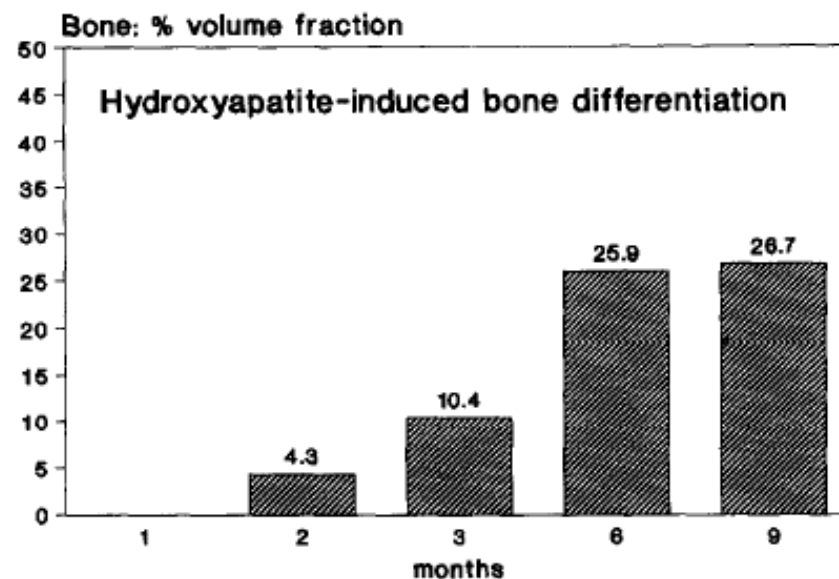


Figure 3 Osteoinduction in coral-derived porous hydroxyapatite after intramuscular implantation in baboons and harvesting at 1, 2, 3, 6 and 9 months. Cumulative means (in %) of histomorphometrical data from Refs 6–9.



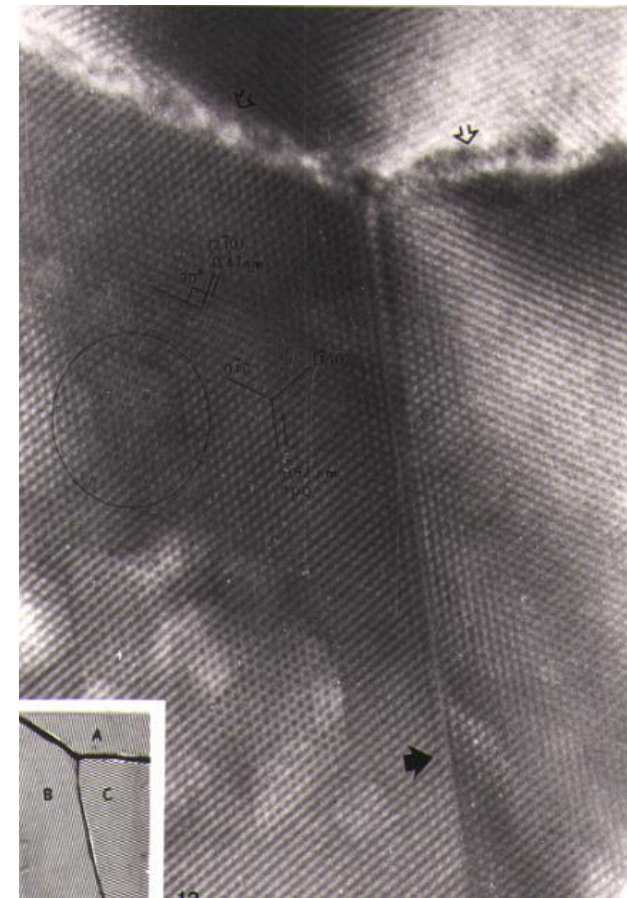
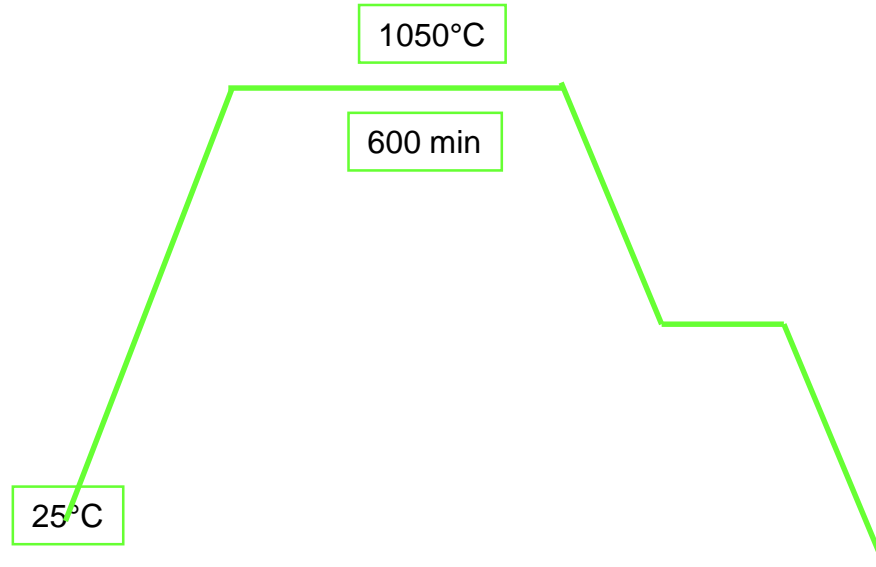
Biomaterials 17 (1996) 31–35
© 1995 Elsevier Science Limited
Printed in Great Britain. All rights reserved
0142-9612/96/\$15.00

Osteoinduction in porous hydroxyapatite implanted in heterotopic sites of different animal models

Ugo Ripamonti

Medical Research Council/University of the Witwatersrand, Bone Research Laboratory, Medical School, 7 York Road, Parktown, 2193 Johannesburg, South Africa

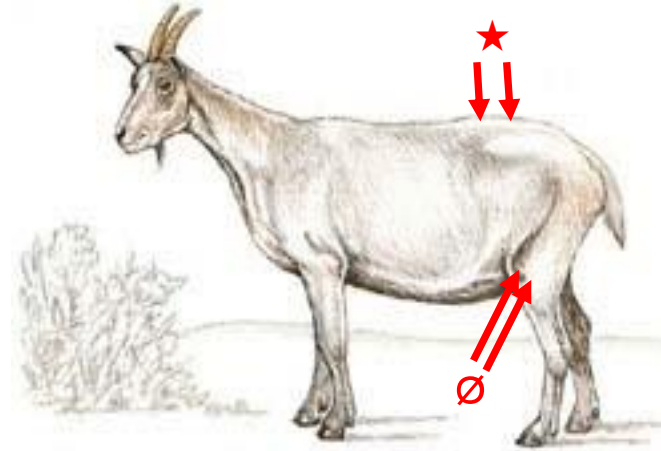
Frittage



Animal model:

2 goats for 6 weeks

7 goats for 12 weeks



Implant:

Teflon cylinder

2 mm thick walls

(Femoral site)



BCP Granules 1-2 mm

1050°C, 1125°C, 1200°C



autologous bone chips

BCP ceramics micro porosity

3 sintering
Methods

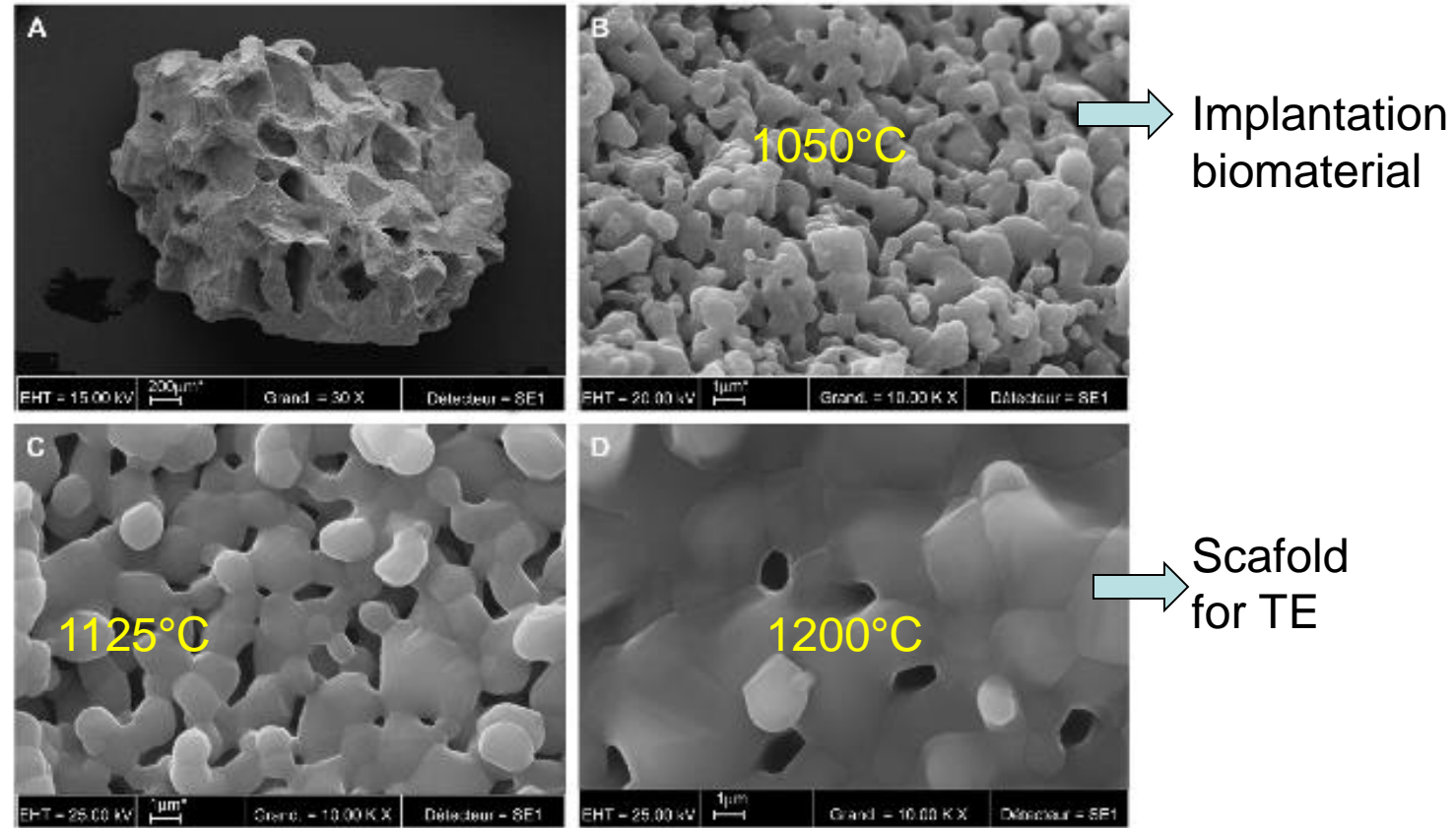


Fig. 1. SEM micrographs of the BCP granules sintered at different temperatures. (A) General shape of a BCP granule. (B)–(D) High magnification images showing the microstructure of BCP granules sintered at 1050, 1125 and 1200 °C, respectively.



Available online at www.sciencedirect.com



Biomaterials 29 (2008) 1177–1188

Biomaterials

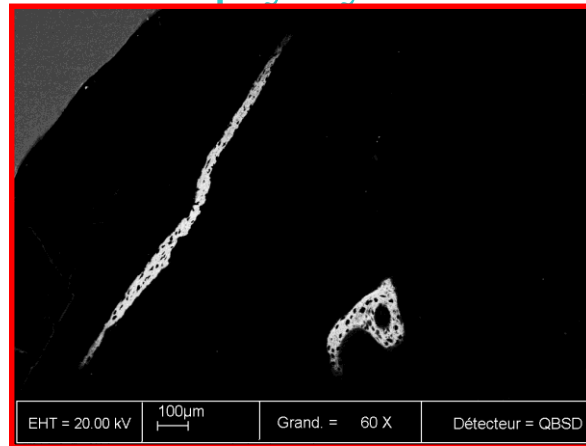
www.elsevier.com/locate/biomaterials

Osteogenicity of biphasic calcium phosphate ceramics
and bone autograft in a goat model

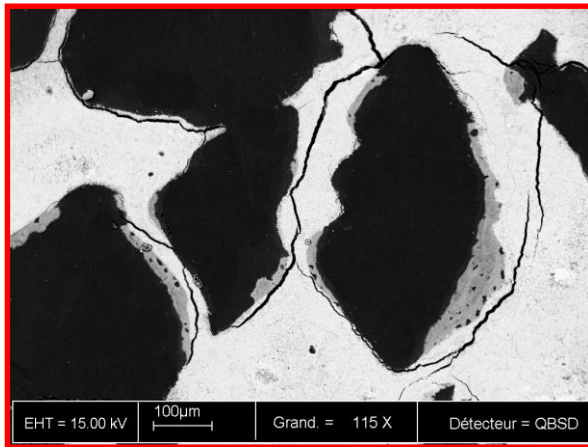
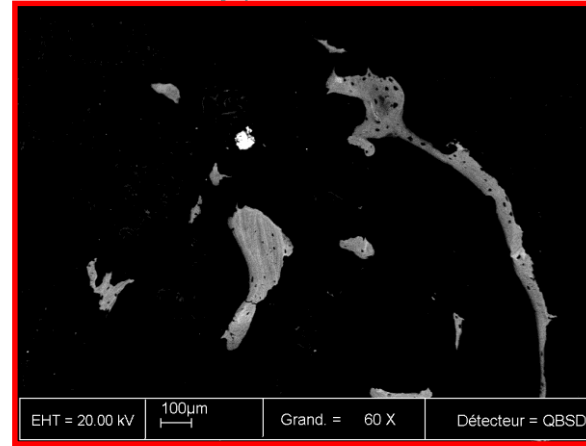
Borhane H. Fella^a, Olivier Gauthier^b, Pierre Weiss^a, Daniel Chappard^c, Pierre Layrolle^{a,*}

Results : Microstructure

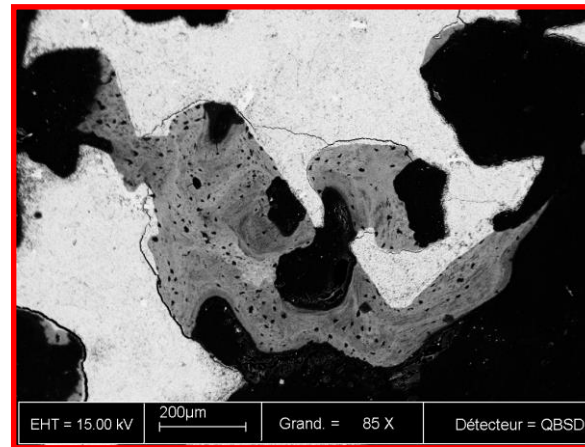
Empty cylinder



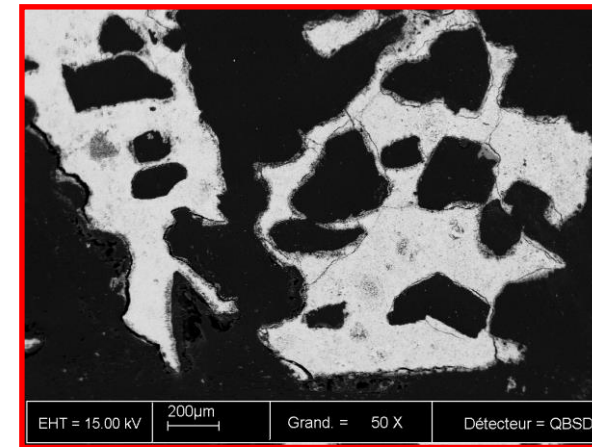
Autologous bone



BCP 1050°C



BCP 1125°C



BCP 1200°C

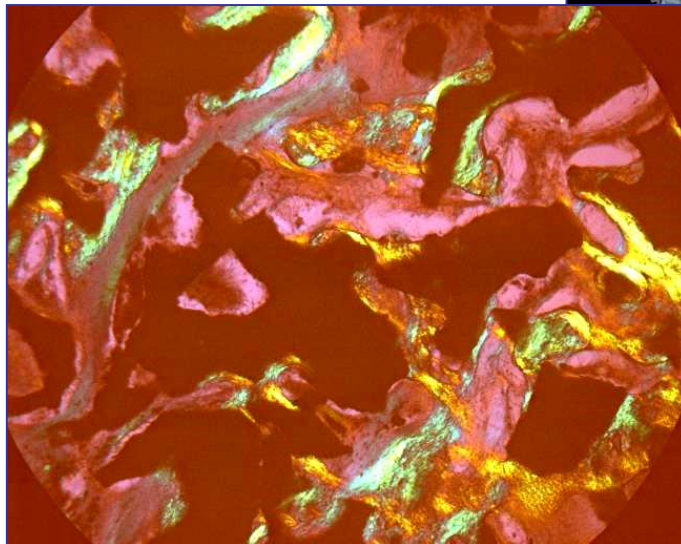
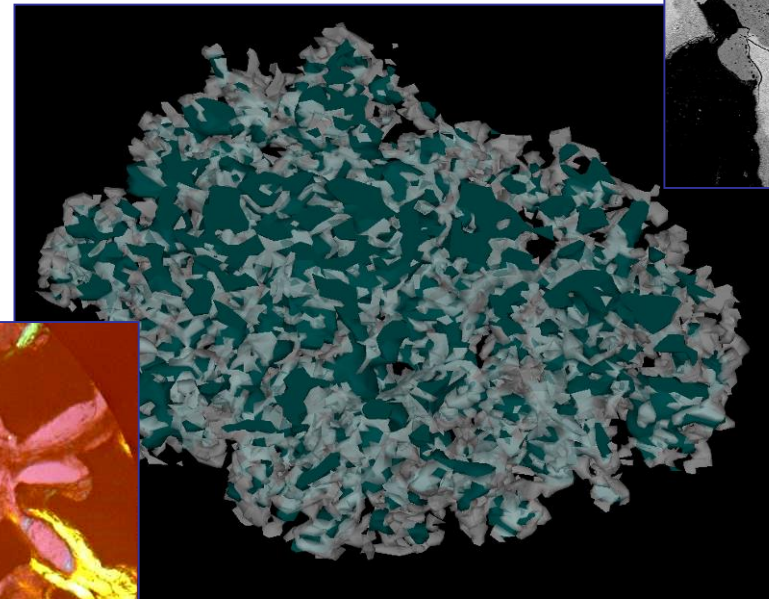
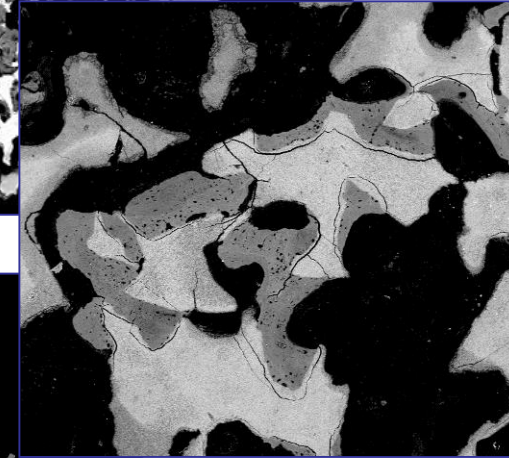
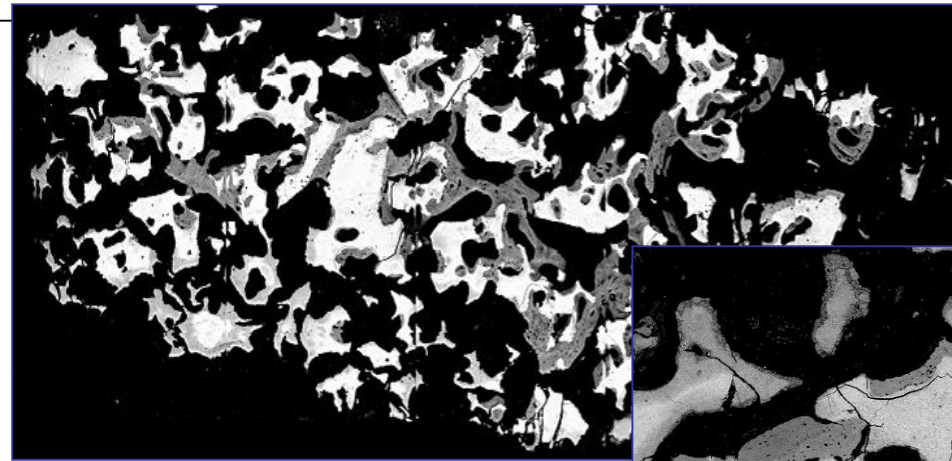
Femurs 12 weeks

Osteoinduction

Muscular area

Granules MBCP 1-2mm

6 months in goat



Micropores effect

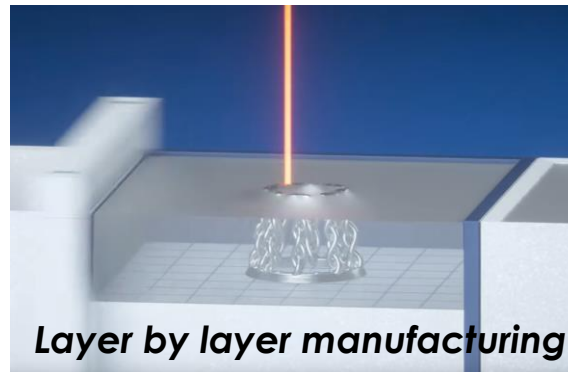
Calcif.Tiss.Res. 1990

Bone 2005

Biofabrication : Additive manufacturing

Additive Manufacturing

- “Bottom-up” procedure



Additive Manufacturing Technologies



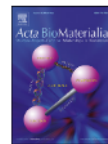


[Acta Biomaterialia 121 \(2021\) 1–28](#)

Contents lists available at [ScienceDirect](#)

Acta Biomaterialia

journal homepage: www.elsevier.com/locate/actbio



Review article

Additive manufacturing pertaining to bone: Hopes, reality and future challenges for clinical applications

Baptiste Charbonnier*, Mikhael Hadida, David Marchat

Mines Saint-Etienne, Université de Lyon, Université Jean Monnet, INSERM, U 1059 Sainbiose, 158, cours Fauriel, CS 62362, 42023 Saint-Etienne Cedex 2, France

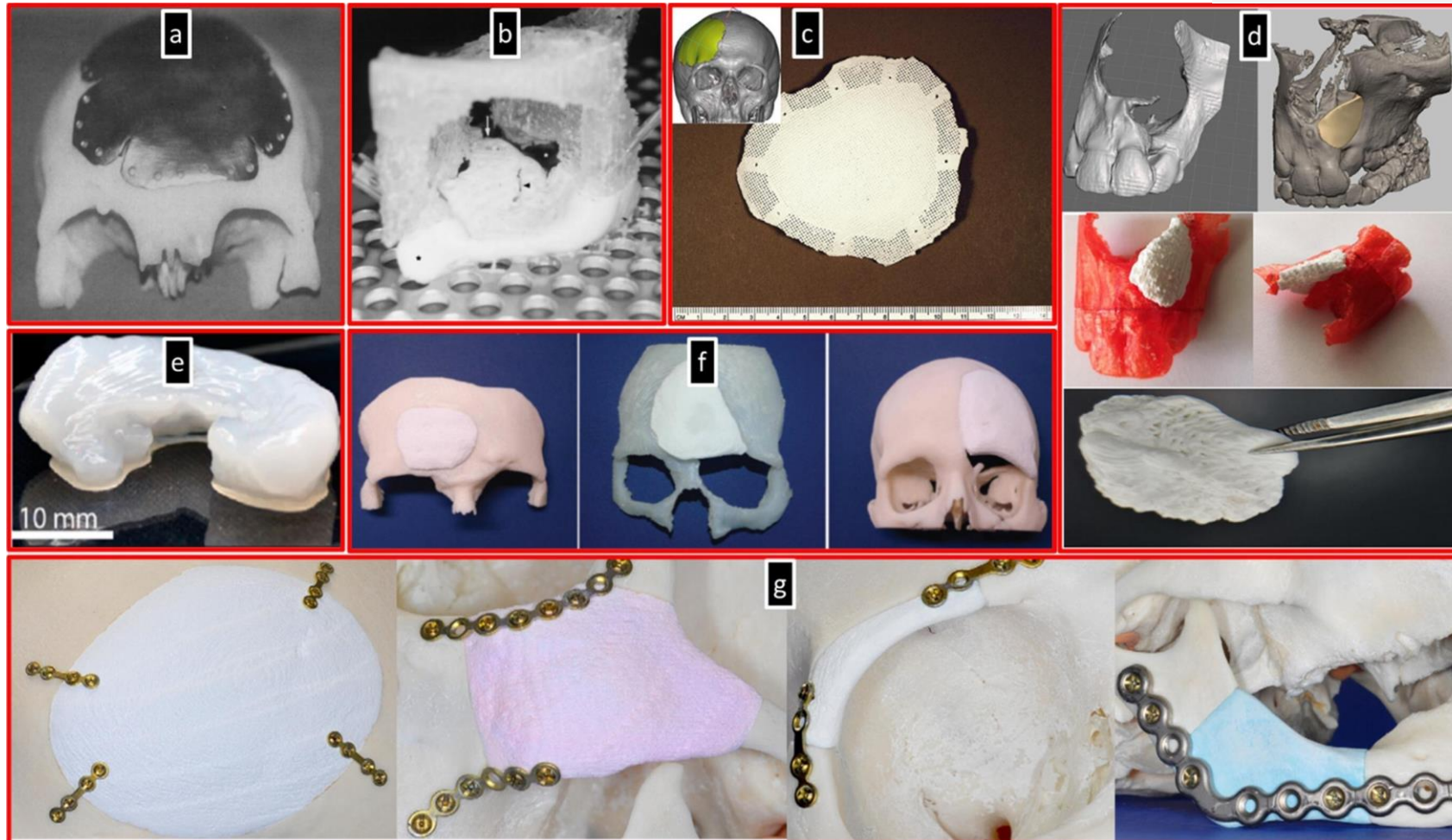


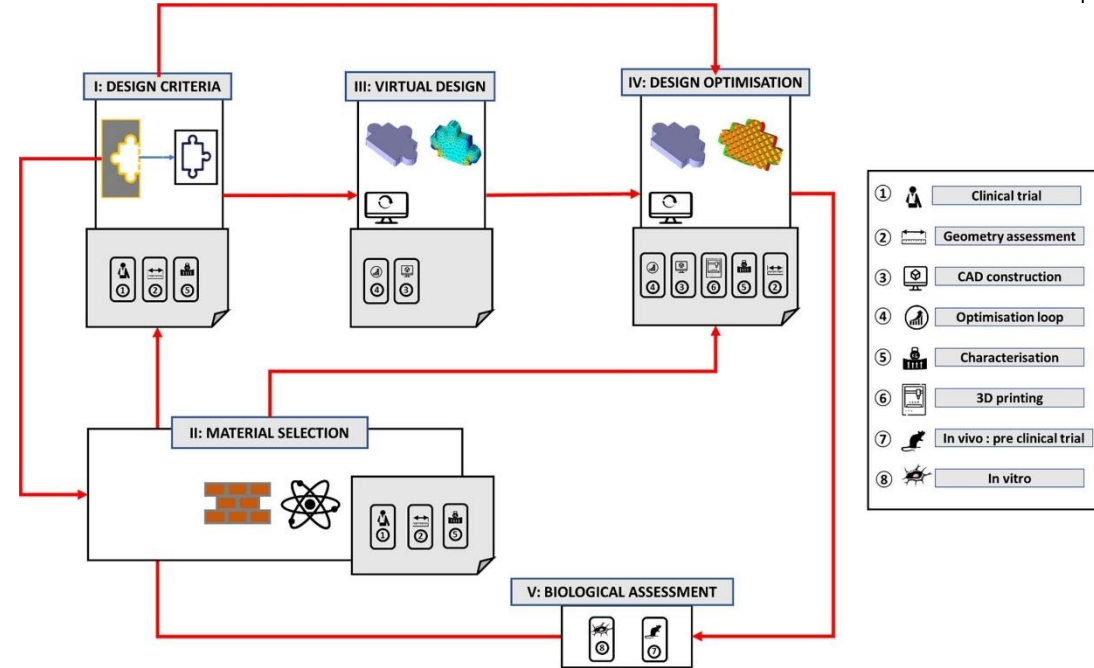
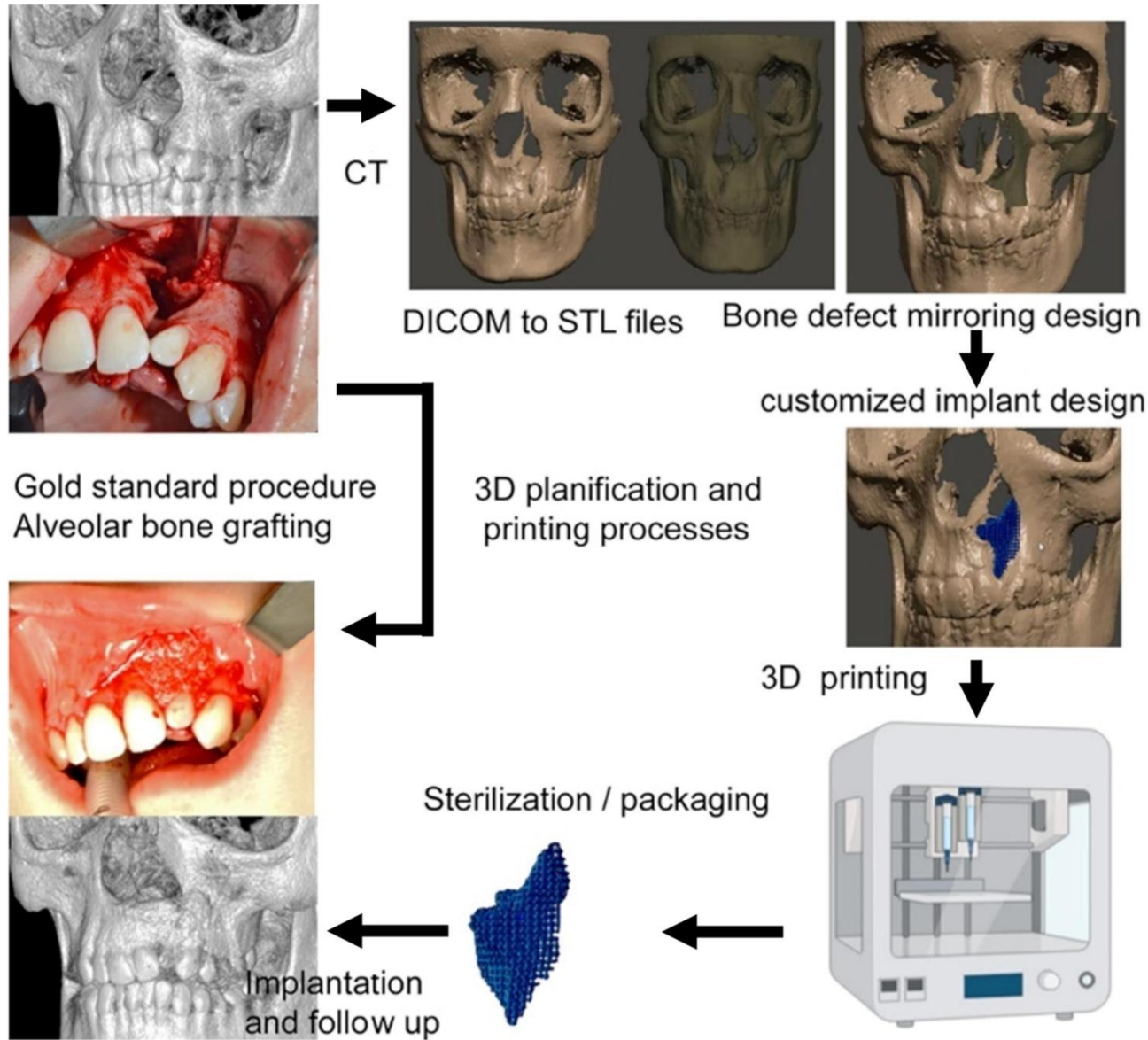


Additive manufacturing of biomaterials for bone tissue engineering – A critical review of the state of the art and new concepts

Marie-Michèle Germaini^a, Sofiane Belhabib^b, Sofiane Guessasma^c, Rémi Deterre^b, Pierre Corre^a, Pierre Weiss^{a,*}

Various biomaterial/AM process couples found in the literature for printing personalized implants. a) titanium/SLS process b) Hydroxyapatite/SLA process c) Hydroxyapatite/SLA process d) Calcium phosphate cement/extrusion-based process e) Hydrogel of alginate/extrusion-based process f) Calcium phosphate cement/powder bed process g) Calcium phosphate cement/powder bed process





Progress in Materials Science 130 (2022) 100963

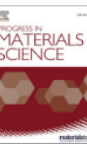


ELSEVIER

Contents lists available at ScienceDirect

Progress in Materials Science

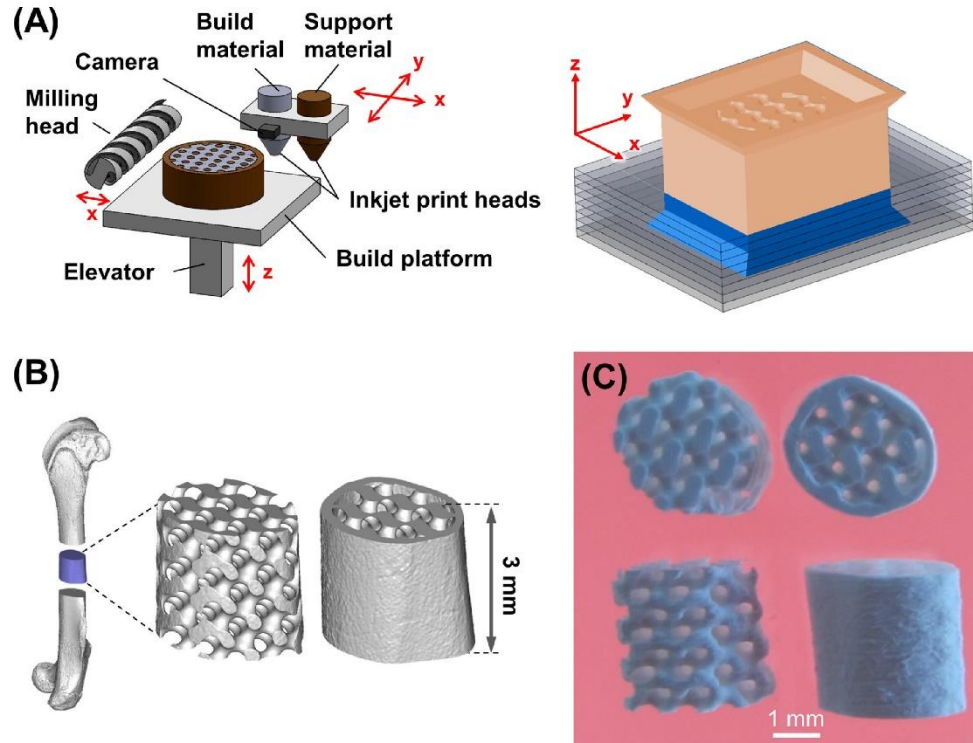
journal homepage: www.elsevier.com/locate/pmatsci



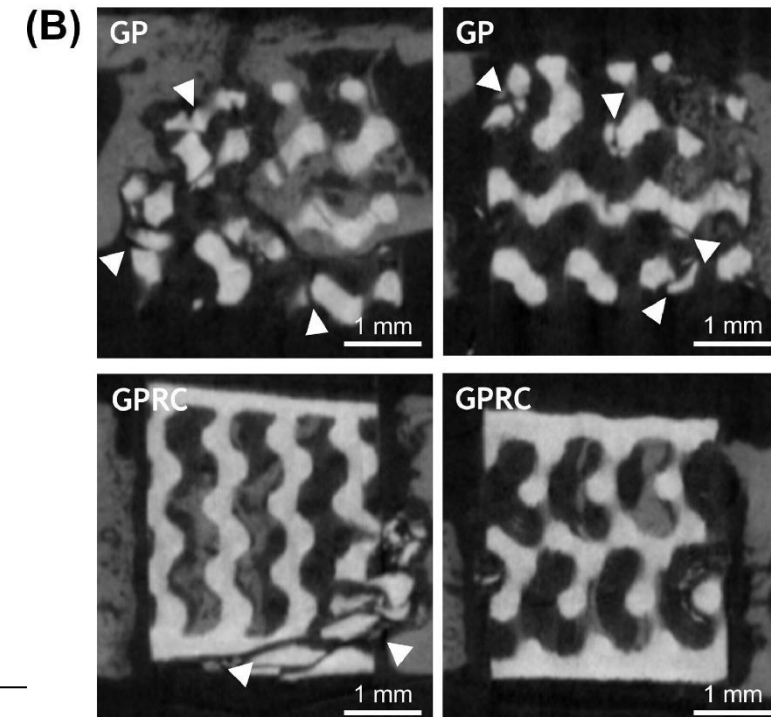
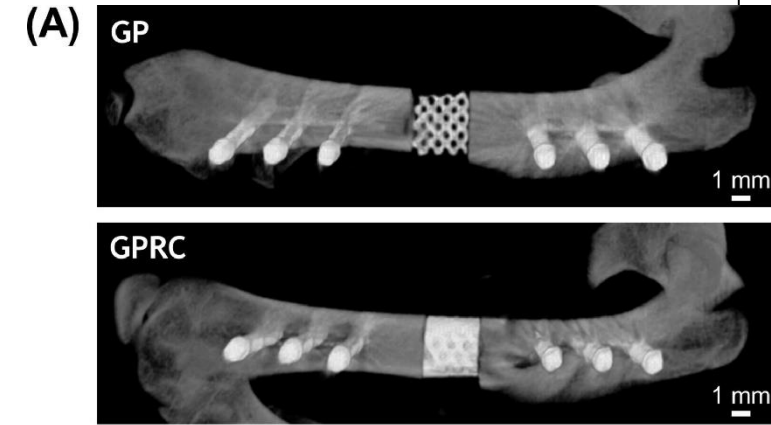
Additive manufacturing of biomaterials for bone tissue engineering – A critical review of the state of the art and new concepts

Marie-Michèle Germaini^a, Sofiane Belhabib^b, Sofiane Guessasma^c, Rémi Deterre^b, Pierre Corre^a, Pierre Weiss^{a,*}

3 D microporous controlled structures



- Printing of 3D Wax mold
- CaP slurry before Sintering
- Gyroid porosity structure without GP or with cortical like outer shell GPRC
- Implanted in a rat femur (8 weeks)

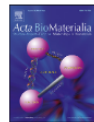


Acta Biomaterialia 109 (2020) 254–266

Contents lists available at ScienceDirect

Acta Biomaterialia

journal homepage: www.elsevier.com/locate/actbio



Full length article

Custom-made macroporous bioceramic implants based on triply-periodic minimal surfaces for bone defects in load-bearing sites



Baptiste Charbonnier^a, Mathieu Manassero^{b,c}, Marianne Bourguignon^{b,c}, Adeline Decambon^{b,c}, Hanane El-Hafci^{b,c}, Claire Morin^a, Diego Leon^a, Morad Bensidoum^{b,c}, Simon Corsia^{b,c}, Hervé Petite^{b,c}, David Marchat^a, Esther Potier^{b,c,*}

Tailored Three-Dimensionally Printed Triply Periodic Calcium Phosphate Implants: A Preclinical Study for Craniofacial Bone Repair

Arnaud Paré,^{†,‡,§,||,◆} Baptiste Charbonnier,^{⊥,◆} Pierre Tournier,^{†,||} Caroline Vignes,[†] Joëlle Veziers,[†] Julie Lesoeur,[†] Boris Laure,^{‡,§} Hélios Bertin,^{||,#} Gonzague De Pinieux,^{§,▽} Grégory Cherrier,^{§,▽} Jérôme Guicheux,^{†,||} Olivier Gauthier,^{†,||,○} Pierre Corre,^{†,||,#} David Marchat,^{⊥,◆} and Pierre Weiss^{*,†,||,◆}

ACS Biomaterials Science & Engineering

Article

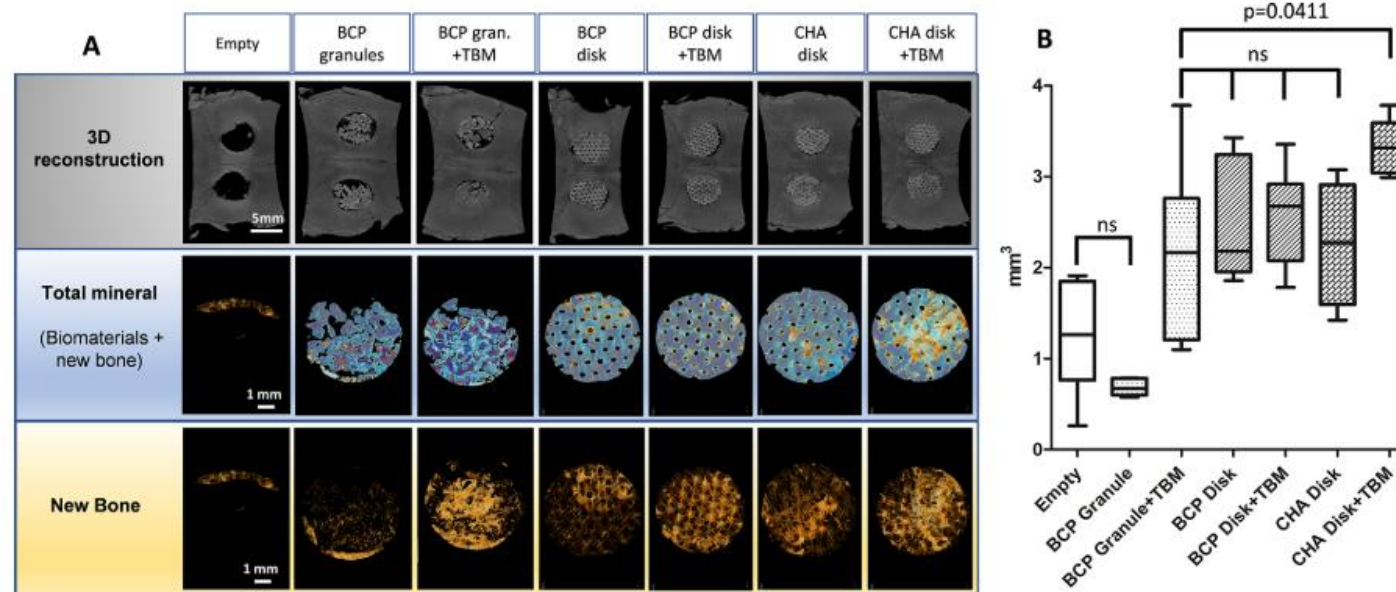
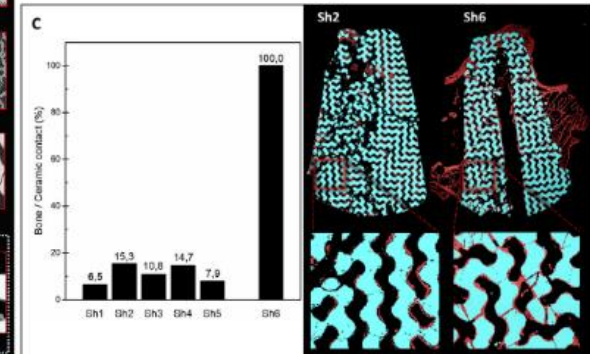
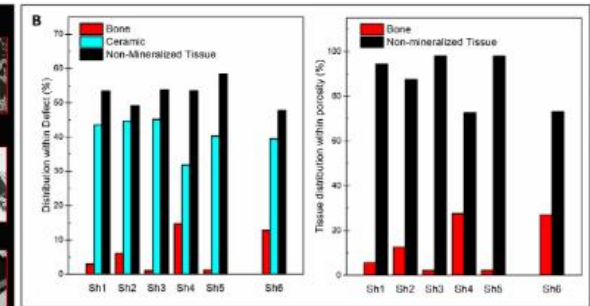
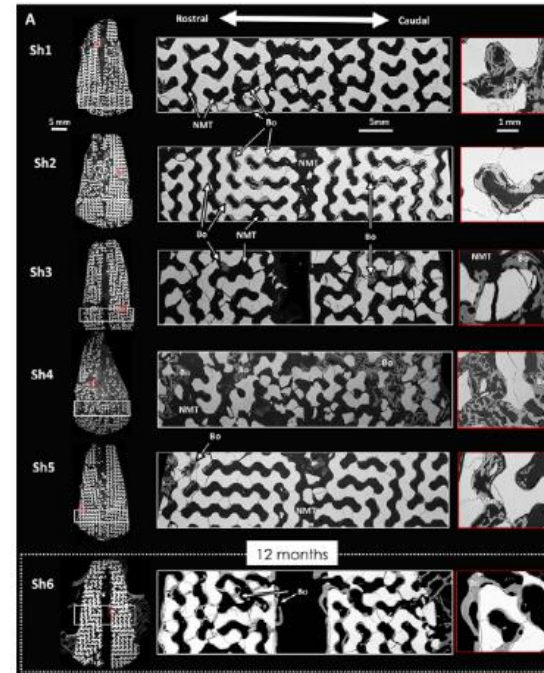
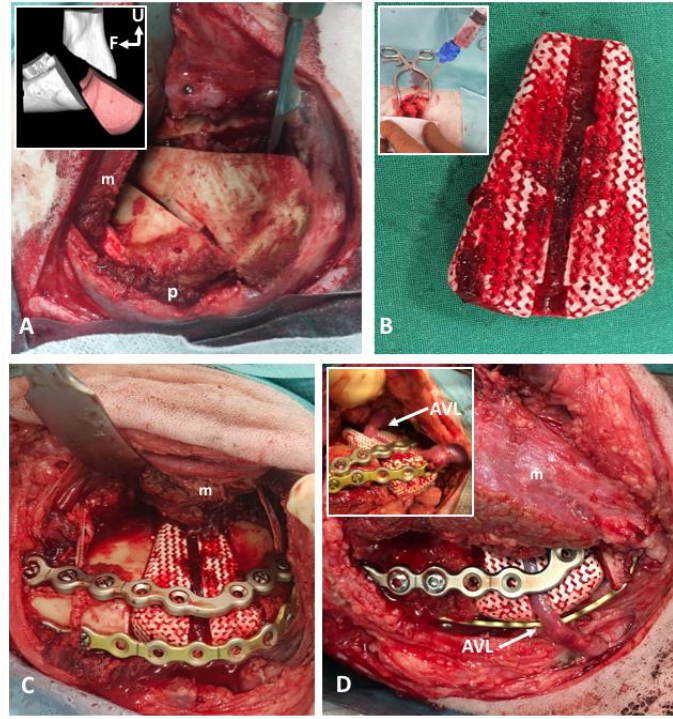
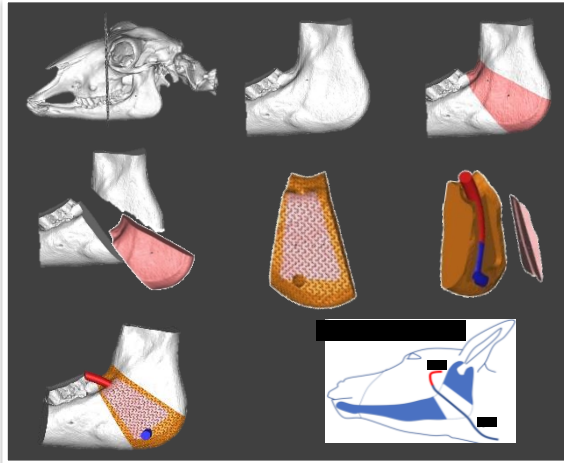


Figure 4. Micro-CT analysis of the critical-sized craniofacial bone defect (CSD) reconstruction. (A) Images of the CSD reconstructions at seven weeks showing calvarial 3D reconstruction, biomaterials + new bone, and newly formed bone alone. CSD repair with BCP granules ± TBM systemically had biomaterial loss and calvarial holes. (B) Graph showing the quantitative analysis of bone volume (BV, mm³) in the region of interest. Empty defect (negative control) and BCP granule groups had the lowest rate of bone formation compared to others ($p < 0.05$). There was no statistical difference between BCP granules + TBM (positive control) and macroporous disks alone, while the CHA + TBM group had significantly higher bone formation than that of BCP granule + TBM ($p = 0.0441$).

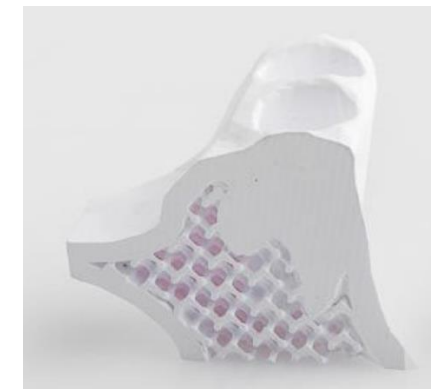
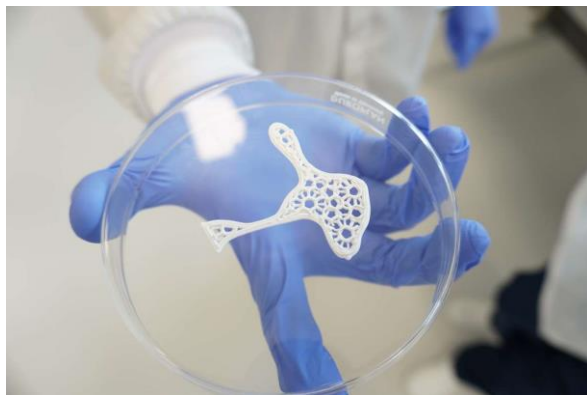
AXIALLY VASCULARIZED CUSTOM-MADE MACROPOROUS BIOCERAMIC FOR RECONSTRUCTION OF SEGMENTAL MANDIBULAR DEFECT: A PRECLINICAL STUDY IN SHEEP

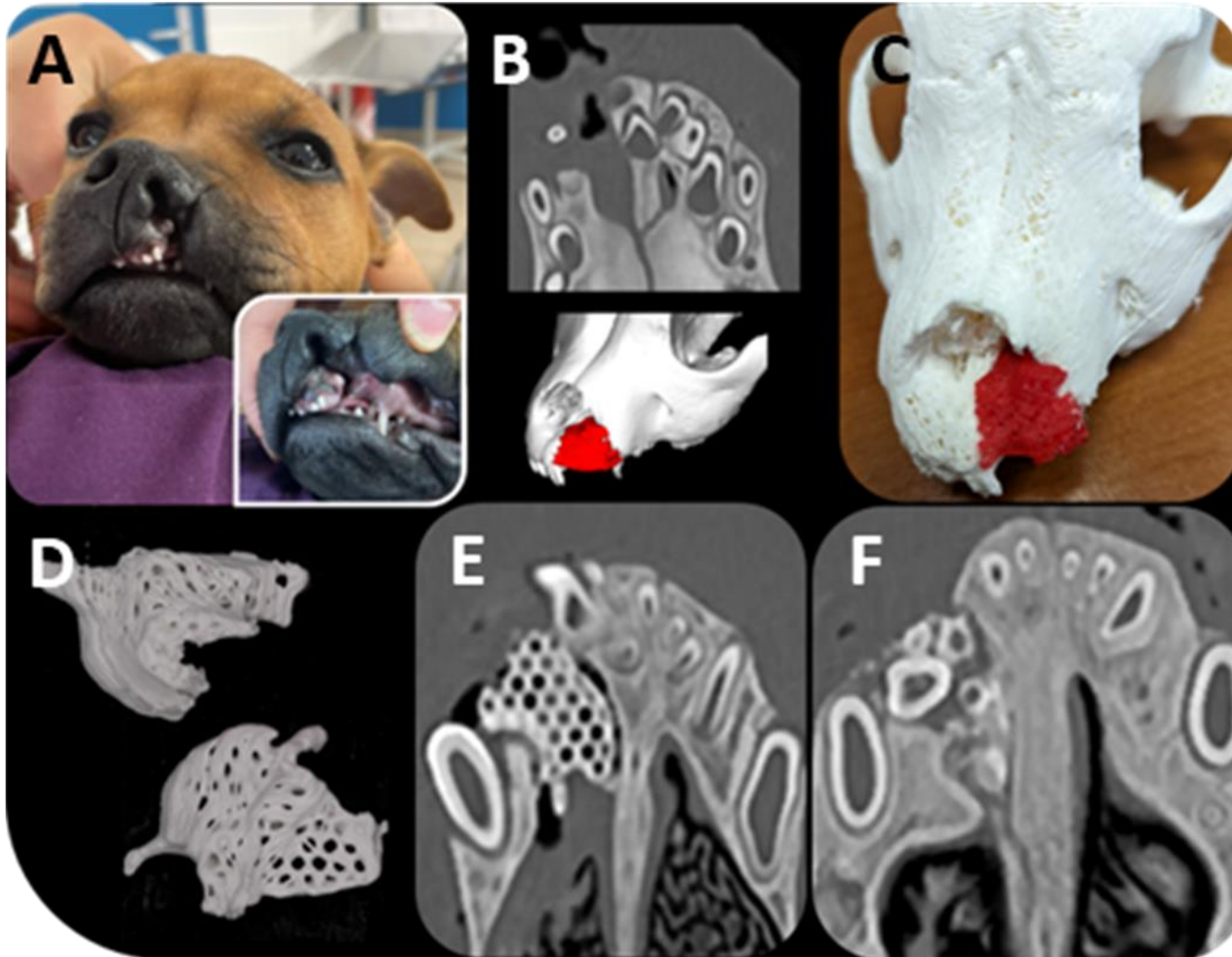


Without polymer or hydrogel **cement is a fragile material**

Additive Manufacturing

- Hopes for a very soon application
 - More and more startups & manufacturers
 - FDA/C€ certification





Traitement personnalisé d'un patient chien comportant (A) une fente labiopalatine unilatérale, (B) le scanner préopératoire 2 semaines avant intervention et la conception d'un implant sur mesure (C) la simulation de la chirurgie via modèles 3D polymères (D) l'implant sur mesure conçu en 2 parties pour faciliter son insertion dans le défaut (E) le scanner post-op après la chirurgie reconstructrice et (F) le scanner à 3 mois post-op, montrant une formation osseuse pouvant supporter l'éruption de dents définitives

→ For tomorrow